Instructions for Post-Transplant Essential Data (Post-TED) Form (Revision 6)

This section of the CIBMTR Forms Instruction Manual is intended to be a resource for completing the Post-Transplant Essential Data (Post-TED) Form (Revision 6).

**Post-Transplant Essential Data (Post-TED)**

Transplant centers participating in the CIBMTR must submit a Post-TED Form for recipients who meet any of the following criteria:

- Recipient receives a transplant at a United States center designated as a *TED-only* center.
- Recipient receives a transplant at a United States center designated as Comprehensive Report Form center and has been assigned to the TED track by the Form Selection Algorithm.
- Recipient receives an allogeneic transplant at a United States center designated as Comprehensive Report Form center but has not consented to participate in research.
- Recipient receives a transplant at an international center, has consented to participate in research, and has been assigned to the TED track by the Form Selection Algorithm.

The Post-TED fulfills the requirements of the Stem Cell Therapeutic Outcomes Database for recipients meeting any of the above criteria. For more information regarding the SCTOD, see General Instructions, Stem Cell Therapeutics Outcomes Database.

For more information, including information on the TED and Comprehensive Report Form Selection Algorithm, see Section 1 in the Center Reference Guide.

The Post-TED must be completed at the following time points: 100 days, six months, and annually post-HCT. These forms should be completed as closely to these time points as possible. The structure of the TED Forms is such that each form should fit on a timeline with distinct start and stop dates that do not overlap any other forms, except in the case of a subsequent HCT.
Subsequent HCT:
If a recipient receives a subsequent HCT between Post-TED time points (100 day, six months, annually), the TED form sequence will start over again with another Pre-TED.

However, if the recipient receives an autologous HCT as a result of a poor graft or graft failure, the TED form sequence will not start over again. Generally, this type of infusion (autologous rescue) is used to treat the recipient’s poor graft response, rather than to treat the recipient’s disease.

Contact the CIBMTR Customer Service Center if the subsequent Pre-TED does not come due automatically.

Non-Malignant Diseases
If the HCT being reported was given to treat a non-malignant disease, as reported on the Pre-TED Disease Classification (2402) Form, do not complete the following sections of the Post-TED Form:
- Q85 – 107: Disease Assessment at the Time of Best Response to HCT
- Q108 – 112: Post-HCT Therapy
- Q113 – 123: Relapse or Progression Post-HCT
- Q124 – 127: Current Disease Status

Lost to Follow-Up:
Occasionally, centers may lose contact with recipients for a variety of reasons, including the recipient’s moving, changing physicians, or death. If contact with a recipient appears lost, please consider calling the recipient at home or work, sending a letter, communicating with the treating or referring physician, or contacting the hospital billing department. If your center receives documented information that a recipient is alive or dead, the form can be marked with only the recipient’s survival status using the Survival Form Status tool in FormsNet3SM.

If no documentation exists and several unsuccessful attempts have been made to contact the recipient, they are considered lost to follow-up and the form may be marked...
as such using the Lost to Follow-Up tool in FormsNet3SM for each reporting period in which no contact exists.

Links to Sections of the Form:
Q1 – 6: Survival
Q 7 – 13: Subsequent Transplant
Q14 – 16: Initial ANC Recovery
Q17 – 18: Initial Platelet Recovery
Q19 – 44: Graft-Versus-Host Disease
Q45 – 47: Liver Toxicity Prophylaxis
Q48 – 49: Veno-occlusive disease (VOD) / Sinusoidal obstruction syndrome (SOS)
Q50 – 56: Infection
Q57 – 64: New Malignancy, Lymphoproliferative or Myeloproliferative Disorder
Q65 – 84: Chimerism Studies (Cord Blood Units, Beta Thalassemia, and Sickle Cell Disease Only)
Q85 – 107: Disease Assessment at the Time of Best Response to HCT
Q108 – 112: Post-HCT Therapy
Q113 – 123: Relapse or Progression Post-HCT
Q124 – 127: Current Disease Status

Manual Updates:
Sections of the Forms Instruction Manual are frequently updated. The most recent updates to the manual can be found below. For additional information, select the manual section and review the updated text.

If you need to reference the historical Manual Change History for this form, please click here or reference the retired manual section on the Retired Forms Manuals webpage.

<table>
<thead>
<tr>
<th>Date</th>
<th>Manual Section</th>
<th>Add/Remove/Modify</th>
<th>Description</th>
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</thead>
</table>

**Q1 – 6: Survival**

The date of actual contact with the recipient to determine medical status for this follow-up report is based on a medical evaluation conducted by a clinician with responsibility for the recipient’s care. Report the date of the medical evaluation performed closest to the designated time period of the form (e.g., Day+100, 6 months, or annual follow-up visit). Time windows are provided to guide selection of dates for reporting purposes.
Recipients are not always seen within the time windows used for reporting follow-up dates, and some discretion is therefore required when determining which date to report. If the recipient is not seen within the time windows, report the date closest to the date of contact within reason.

If the Post-TED Form reports a subsequent infusion (transplant or genetically modified cellular therapy), report the date of latest follow-up as the day prior to the start of the preparative regimen / systemic therapy. If no preparative regimen or conditioning / systemic therapy was given, report the day prior to infusion as the date of contact.

**Reporting Latest Follow-up**
When reporting the date of latest follow-up prior to a subsequent HCT, report the date specified above regardless whether there is actual patient contact on the date. This is an exception to standard date of follow-up reporting to ensure all dates are captured within the sequence of forms.

**Reporting the 1-Year Date of Contact**
If this form is being completed for the 1-year reporting period, ensure the reported contact date is ≥ Day 365. Review the 1-Year Date of Contact instructions below for additional information.

**Question 1: Date of actual contact with the recipient to determine medical status for this follow-up report**
Enter the date of actual contact with recipient to determine medical status for this follow-up report. Acceptable evaluations include those from the transplant center, referring physician, or other physician currently assuming responsibility for the recipient’s care. If an evaluation was not performed at Day+100, at 6 months, or on the HCT anniversary, choose the date of the visit closest to the actual time point.

If the recipient has not been seen by a clinician during the reporting period but the survival status is known, complete the Survival Tool reference in the CIBMTR Data Management Guide, found here.

In general, the date of contact should be reported as close to the 100 day, 6 month, or annual anniversary to transplant as possible. Report the date of actual contact with the recipient to evaluate medical status for the reporting period. In the absence of contact with a clinician, other types of contact may include a documented phone call with the recipient, a laboratory evaluation, or any other documented recipient interaction on the date reported. If there was no contact on the exact time point, choose the date of contact closest to the actual time point. Below, the guidelines show an ideal approximate range for reporting each post-transplant time point:

<table>
<thead>
<tr>
<th>Time Point</th>
<th>Approximate Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>100 days</td>
<td>+/- 15 days (Day 85-115)</td>
</tr>
</tbody>
</table>
Recipients are not always seen within the approximate ranges and some discretion is required when determining the date of contact to report. In that case, report the date closest to the date of contact within reason. The examples below assume that efforts were undertaken to retrieve outside medical records from the primary care provider, but no documentation was received.

**Example 1. The 100 day date of contact doesn’t fall within the ideal approximate range.**

The autologous recipient was transplanted on 1/1/13 and is seen regularly until 3/1/13. After that, the recipient was referred home and not seen again until 7/1/13 for a restaging exam and 7/5/13 for a meeting to discuss the results.

What to report:
- **100 Day Date of Contact:** 3/1/13 (Since there was no contact closer to the ideal date of 4/11/13, this date is acceptable)
- **6 Month Date of Contact:** 7/5/13 (note the latest disease assessment would likely be reported as 7/1/13)

**Example 2. The 100 day date of contact doesn’t fall within the ideal approximate range and the recipient wasn’t seen again until 1 year post-HCT.**

The autologous recipient was transplanted on 1/1/12 and is seen regularly until 3/1/12. After that, the recipient was referred home and not seen again until 1/1/13 for a restaging exam and 1/4/13 for a meeting to discuss the results.

What to report:
- **100 Day Date of Contact:** 3/1/13 (Since there was no contact closer to the ideal date of 4/11/13, this date is acceptable)
- **6 Month Form:** Mark the form Lost to Follow-Up using the Lost to Follow-Up Tool in FormsNetSM
- **1 Year Date of Contact:** 1/4/13 (note the latest disease assessment would likely be reported as 1/1/13)

Additional information:
- A date of contact should never be used multiple times for the same recipient’s forms.
- For example, 6/1/13 should not be reported for both the 6 month and 1 year form. Instead, determine the best possible date of contact for each reporting
period; if there is not a suitable date of contact for a reporting period, this may indicate that the recipient was lost to follow-up.

- If the recipient has a disease evaluation just after the ideal date of contact, capturing that data on the form may be beneficial.

- For example, if the recipient’s 90 day restaging exam was delayed until day 115 and the physician had contact with the recipient on day 117, the restaging exams can be reported as the latest disease assessment and day 117 would be the ideal date of contact, even though it is just slightly after the ideal approximate range for the date of contact.

**Date of Contact & Death**

In the case of recipient death, the date of death should be reported as the date of contact regardless of the time until the ideal date of contact. The date of death should be reported no matter where the death took place (inpatient at the transplant facility, at an outside hospital, in a hospice setting, or within the recipient’s home).

If the death occurred at an outside location and records of death are not available, the dictated date of death within a physician note may be reported. If the progress notes detailing the circumstances of death are available, request these records. These records are useful for completing required follow-up data fields and the cause of death data fields on this form. If the exact date of death is not known, use the processed described for reporting partial or unknown dates, see General Instructions, **General Guidelines for Completing Forms**.

**Example 3.** *The recipient has died before their 6 month anniversary.*

The recipient is transplanted on 1/1/13, was seen regularly through the first 100 days. They had restaging exams on 4/4/13 and was seen on 4/8/13, and then died on 5/13/13 in the hospital emergency room.

What to report:

- **100 Day Date of Contact:** 4/8/13 (note the latest disease assessment would likely be reported as 4/4/13)
- **6 Month Date of Contact:** 5/13/13 (though the death does not occur within the ideal approximate range for 6 months)

**Example 4.** *The recipient has died after their 6 month anniversary.*

The recipient is transplanted on 1/1/13, was seen regularly through the first 100 days. They had restaging exams on 4/22/13 and was seen on 4/23/13. Based on findings in the restaging exam, the recipient was admitted for additional treatment. The disease was found to be refractory on a 6/25/13 restaging exam, and the recipient was discharged to hospice on 7/8/13. The hospital was notified via telephone that the recipient died on 7/16/13.

What to report:
100 Day Date of Contact: 4/23/13 (note the latest disease assessment would likely be reported as 4/22/13)
6 Month Date of Contact: 7/16/13 (note the latest disease assessment would likely be reported as 6/25/13)

Date of Contact & Subsequent Transplant
If the recipient has a subsequent infusion (HCT or cellular therapy), the date of contact will depend on the type of subsequent infusion. If the subsequent infusion is an HCT or genetically modified cellular therapy (e.g. CAR-T), report the date of contact as the day before the preparative regimen / systemic therapy begins for the subsequent infusion. If no preparative regimen / systemic therapy is given, report the date of contact as the day before the subsequent infusion. In these cases, actual contact on that day is not required, and the day prior to the initiation of the preparative regimen (or infusion if no preparative regimen / systemic therapy) should be reported. This allows every day to be covered by a reporting period but prevents overlap between infusion events. If the subsequent infusion is a non-genetically modified (e.g. DLI) cellular therapy infusion, report the date of contact as appropriate to the reporting period.

Example 5. The recipient had a 2nd transplant with a preparative regimen.
The recipient has their first transplant on 1/1/13 and a planned second transplant on 2/1/13. The recipient was admitted on and received their first dose of chemotherapy for the preparative regimen for HCT #2 on 1/28/13.

What to report:
100 Day Date of Contact: 1/27/13 (regardless of actual contact on that date)

Example 6. The recipient had a subsequent transplant without a preparative regimen.
Following their first transplant on 1/1/13, a recipient with SCID required a subsequent allogeneic transplant due to poor graft function. The recipient has remained inpatient following the first transplant. The physician planned the second transplant for 5/31/13, and proceeded without a preparative regimen.

What to report:
100 Day Date of Contact: 4/11/13 (+/- 15 days)
6 Month Date of Contact: 5/30/13

Example 7. The recipient had a subsequent genetically modified cellular therapy with lymphodepleting therapy administered prior to infusion.
The recipient has their first transplant on 2/1/15 and a genetically modified (e.g. CAR-T) cellular therapy infusion on 3/1/15. The recipient was admitted on and received their first dose of lymphodepleting therapy 2/28/15.
What to report:

100 Day Date of Contact: 2/27/15 (regardless of actual contact on that date). Both HCT and cellular therapy forms will be completed but all applicable HCT follow-up forms will be reset to the new event date (i.e., Forms 4100+2450). See Subsequent Infusions – Updates to Follow-Up Reporting in the Data Management Manual for more information on combined follow up.

**Example 8. The recipient had a subsequent non-genetically modified cellular therapy.**
The recipient has their first transplant on 1/21/15 and a non-genetically modified (e.g. DLI) cellular therapy infusion on 2/15/15. There was no lymphodepleting therapy administered.

What to report:
100 Day Date of Contact: The date of contact reported will be appropriate to the reporting period. Combined follow up will not be applied, a single F4100 is required, then HCT reporting continues.

1-Year Date of Contact
When reporting the date of contact for the 1-year reporting period, if the recipient is alive, report a contact date on or after Day 365. The date of contact should not be reported prior to Day 365 for the 1-year reporting period. This ensures the recipient is included in the numerator for the transplant center’s Center Specific Analysis (CSA).

**Example 9. A recipient is evaluated before and after Day 365 but not on Day 365**
The recipient had an allogeneic transplant on 1/5/13 and is seen regularly until 6/20/13. After that, the recipient was referred home and not seen again until 1/1/14 for a restaging exam and again on 1/15/14 to review the results. Day 365 is 1/5/14.

What to report:
1-Year Date of Contact: 1/15/14 (since this date is ≥ Day 365)

**Example 10. A recipient is evaluated before and after Day 365 but not on Day 365**
The recipient is transplanted on 2/28/19 and seen regularly until 8/28/19. The next visit is on 2/20/20 for blood work and the lab results are phoned to the recipient on 2/21/20. The recipient was not evaluated again until 4/1/20. Day 365 is 2/28/20.

What to report:
1-Year Date of Contact: 4/1/20
For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

**Question 2: Specify the recipient’s survival status at the date of last contact**
Indicate the clinical status of the recipient on the date of actual contact for follow-up evaluation.

**Question 3: Primary cause of death**
Report the underlying cause of death. Do not report the mode of death, such as cardiopulmonary arrest. According to the Centers for Disease Control and Prevention, National Center for Health Statistics, the underlying cause of death is “the disease or injury that initiated the chain of events that led directly or inevitably to death.”

Report only one primary cause of death; see the Cause of Death Codes section of the Forms Instructions Manual for more details regarding cause of death. If the recipient has recurrent/persistent/progressive disease at the time of death, consider if the disease was the primary cause of death or a contributing cause of death. It should not be assumed that the presence of disease indicates that the disease was the primary cause of death.

**Aplastic Anemia**
If the recipient received an HCT for aplastic anemia, and the primary cause of death is attributed to relapse/recurrence of disease, report “HCT related causes” and select “Rejection/Poor graft function” as the cause of death.

If the primary cause of death is unclear, consult with a physician for their best medical opinion.

**Question 4: Specify**
Specify the details for primary cause of death requiring “other” specification. Options which require additional specification include Other infection, Other pulmonary syndrome, Multiple organ failure, Other organ failure, Other hemorrhage, Other vascular, and Other cause. Information reported in the specify field must pertain to the option selected (e.g., an infectious cause of death should be specified for “Other infection”).

**Question 5: Contributing cause of death:**
Report any additional causes of death. All contributing causes of death are important for analysis of transplant outcomes. Refer to the Cause of Death Codes section of the Forms Instructions Manual for more details regarding cause of death. If there were multiple contributing causes of death, enable an additional instance to report additional causes.

**Question 6: Specify**
Specify the details for contributing cause of death requiring “other” specification. Options which require additional specification include Other infection, Other pulmonary syndrome, Multiple organ failure, Other organ failure, Other hemorrhage, Other vascular, and Other cause. Information reported in the specify field must pertain to the option selected (e.g., an infectious cause of death should be specified for “Other infection”).

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<tr>
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<th>Reasoning (If applicable)</th>
</tr>
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Q7 – 13: Subsequent Transplant

Question 7: Did the recipient receive a subsequent HCT since the date of last report?
Indicate whether the recipient received a second (or third, etc.) hematopoietic stem cell infusion. Hematopoietic stem cells are defined as mobilized peripheral blood stem cells, bone marrow, or cord blood. The source of the hematopoietic stem cells may be allogeneic unrelated, allogeneic related, or autologous. For more information on how to distinguish infusion types (example: HCT versus DCI), see Appendix D.

If the recipient has received a subsequent HCT since the date of the last report, ensure the date of actual contact reported in question 1 is the date immediately prior to the start of the preparative regimen for the subsequent HCT. If no preparative regimen was given, report the date prior to infusion.

Question 8: Date of subsequent HCT
Report the planned or actual date of the subsequent HCT infusion. If the planned date is reported and changes, this field will need to be updated to reflect the actual date of subsequent HCT infusion. If multiple days of infusion are planned, report the first.

Questions 9 – 10: What was the indication for subsequent HCT?
Indicate the reason for the subsequent HCT (check only one).

- **Graft failure / insufficient hematopoietic recovery**: Additional stem cells are required because there wasn’t any ANC recovery following HCT (primary graft failure), the hematopoietic recovery indefinitely declined after the initial hematopoietic recovery or hematopoietic recovery (secondary graft failure), or hematopoietic recovery was deemed insufficient or too slow for survival following previous high-dose therapy and HCT. If autologous cells are infused
for this reason, this is considered autologous rescue; in this case, reporting will continue under the prior HCT date and a new Pre-TED form is not required.

- **Persistent primary disease.** Additional stem cells are required because of the persistent presence of disease pre and post-transplant (i.e., complete remission was never achieved following the previous transplant).

- **Recurrent primary disease:** Additional stem cells are required because of relapsed primary disease (i.e., complete remission was achieved pre or post-transplant, but the disease relapsed following the previous transplant).

- **Planned subsequent HCT, per protocol:** Additional stem cells are given as defined by the protocol for a subsequent transplant/infusion. This transplant is not based upon recovery, disease status, or any other assessment.

- **New malignancy (including PTLD and EBV lymphoma):** Additional stem cells are required because the recipient has developed a new malignancy. This does not include a transformation or progression of the original malignancy for which the recipient was transplanted (refer to the New Malignancy, Lymphoproliferative or Myeloproliferative Disorder section for more information). If **New malignancy** is selected, also complete the New Malignancy, Lymphoproliferative or Myeloproliferative Disorder section of the form.

- **Insufficient chimerism:** In the case of a stable, mixed donor chimerism, the infusion of additional cells (usually lymphocytes and not mobilized stem cells) may be classified as a DCI. However, in the case of declining chimerism – when the percentage of donor cells is sequentially decreasing on several studies, indicating possible impending graft failure (donor chimerism has often fallen below 30-50%) – additional stem cells may be required. Verify with the transplant physician that the cells given should be reported as a subsequent transplant.

- **Other:** If additional stem cells are given for a reason other than the options listed, select **Other** and specify in question 10.

**Question 11: Source of HSCs (check all that apply)**
Report the stem cell source(s) of the recipient’s subsequent HCT. All allogeneic sources, and autologous sources with indication other than “graft failure / insufficient hematopoietic recovery”, will require another Pre-TED form to be completed for the subsequent HCT.

**Question 12: Has the recipient received a cellular therapy since the date of last report? (e.g. CAR-T, DCI)**
If course of cellular therapy carries over an HCT reporting period, and has already been reported on a prior form, do not re-report that course of cellular therapy. For example, if a course of cellular therapy includes three infusions, and the third infusion overlaps from the one year to two-year HCT reporting period, do not report a new cellular therapy on the two year HCT follow up form.

Indicate whether the recipient received a cellular therapy for any reason within the reporting period. The most common type of post-HCT cellular therapy would be a donor cellular infusion (DCI) / donor lymphocyte infusion (DLI). These infusions are not intended to promote hematopoiesis. If the recipient received additional cells due to engraftment issues, or if they received an infusion of unmanipulated CD34+ cellular product (stimulated peripheral blood stem cells, bone marrow, or cord blood), report as a subsequent HCT rather than a cellular therapy. For more information on how to distinguish infusion types (example: HCT versus DCI), see Appendix D.

A DCI is a form of cellular therapy that uses cells from the original donor and is commonly used to create a graft-versus-leukemia / tumor (GVL / GVT) effect. The recipient does not receive a preparative regimen prior to receiving the donor cells because the purpose of a DCI is to activate the immune system rather than repopulate the marrow. The recipient may, however, be given therapy prior to the infusion for the purpose of disease control, or as lymphodepletion. The types of cells used in a DCI include, but are not limited to: lymphocytes, unstimulated peripheral blood mononuclear cells, dendritic cells, and / or mesenchymal cells.

Other forms of cellular therapy may include cytotoxic T-lymphocytes (CTLC) to treat infections or chimeric antigen receptor T-cells (CAR T-cells) to treat persistent, progressive or recurrent disease.

**Question 13: Date of cellular therapy**

Report the date of cellular therapy infusion. If multiple infusions were received in the reporting period, report the earliest. If infusions are continuing from a previous instance of DCI, only report in the period during which

**Section Updates:**

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<th>Description</th>
<th>Reasoning (If applicable)</th>
</tr>
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<tbody>
<tr>
<td>Q14 – 16: Initial ANC Recovery</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Initial ANC Recovery

The Initial ANC Recovery questions can only be completed on the 100-day, 6 month, 1 year, and 2 year follow-up forms. These questions will be skipped for all subsequent reporting periods. Question 16 must be answered on all follow-up forms.

Initial ANC Recovery

Recovery, as reported in this section, does not distinguish between allogeneic engraftment (blood and stem cells of donor origin) and autologous engraftment (blood and stem cells of host origin). To demonstrate engraftment for allogeneic recipients, particularly non-myeloablative or reduced intensity approaches, chimerism tests must be done. These measure the quantity of donor cells relative to the quantity of host (recipient) cells. While ANC usually represents donor cells in allogeneic HCT, it cannot be proven without chimerism studies.

ANC recovery is defined as an absolute neutrophil count (ANC) of $\geq 0.5 \times 10^9/L$ (500/mm³) for three consecutive laboratory values obtained on different days. Date of ANC recovery is the date of the first of three consecutive laboratory values where the ANC is $\geq 0.5 \times 10^9/L$. At some institutions, the laboratory reports display the ANC value once there are sufficient white blood cells to perform a differential count. At other institutions, the laboratory reports do not display the ANC, and it must be calculated from the white blood cell count (WBC) and the percent of segmented and band neutrophils (if the differential was performed on a machine, the percent neutrophils will include both segmented and band neutrophils). If the laboratory report displays an automated ANC value of exactly 0.5, the actual ANC value should be calculated from the manual differential if available. The calculated value from the manual differential will determine ANC recovery. If your institution’s laboratory reports do not display the ANC value, use the following calculation to determine the ANC:

Calculating Absolute Neutrophil Count (ANC)\(^1\)

\[
\frac{\text{% segmented neutrophils}}{\text{+ \% band neutrophils}} = \frac{\text{% neutrophils}}{\text{x white blood cell count/mm}^3} = \text{absolute neutrophil count/mm}^3
\]

Example:

(Divide percentage by 100 to convert to decimal)

\[
\begin{align*}
0.45 \text{ segmented neutrophils} & + 0.05 \text{ band neutrophils} \\
= 0.50 \text{ neutrophils} & \times 1000/mm^3 \text{ white blood cell count} \\
= 500/mm^3 \text{ absolute neutrophil count}
\end{align*}
\]
Traditionally, the definition of ANC recovery required selecting the first date of three consecutive days in which the recipient’s ANC was ≥ 0.5x10⁹/L (500/mm³). For various reasons it may not be possible to obtain daily laboratory values. Under those circumstances, report ANC recovery based upon three consecutive laboratory values (drawn more than a day apart) as long as the ANC remains ≥ 0.5x10⁹/L (500/mm³).

Tracking the date of ANC recovery may not always be straightforward. In some cases, the ANC may fluctuate for a period of time before the recipient fully recovers. In other cases, the ANC may remain above 0.5 × 10⁹/L for several days immediately post-HCT and then fall below 0.5 × 10⁹/L. Do not begin counting ANC values of ≥ 0.5 × 10⁹/L towards recovery until the ANC has dropped to the lowest level (nadir) post-HCT. If the recipient was transplanted using a non-myeloablative (NST) or reduced intensity (RIC) regimen, or was transplanted for an immunodeficiency (e.g., SCID, WAS), the recipient’s ANC may never drop below 0.5 × 10⁹/L. If this is the case, an ANC recovery date will not be reported, and the “never below” option should be chosen. However, if the recipient’s ANC drops below 0.5×10⁹/L for even one day, this should be considered the nadir and “never below” should not be chosen. See the following example for more information regarding tracking the date of ANC recovery.

**Tracking ANC Recovery**

*Transplant Date = May 6*

<table>
<thead>
<tr>
<th>Date</th>
<th>WBC</th>
<th>%Neutrophils</th>
<th>ANC</th>
</tr>
</thead>
<tbody>
<tr>
<td>May 7</td>
<td>900</td>
<td>0.6</td>
<td>540</td>
</tr>
<tr>
<td>May 8</td>
<td>850</td>
<td>0.59</td>
<td>502</td>
</tr>
<tr>
<td>May 9</td>
<td>720</td>
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</tr>
<tr>
<td>May 10</td>
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</tr>
<tr>
<td>May 11</td>
<td>15</td>
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<td>—</td>
</tr>
<tr>
<td>May 12</td>
<td>30</td>
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<tr>
<td>May 14</td>
<td>250</td>
<td>0.4</td>
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</tr>
<tr>
<td>May 15</td>
<td>800</td>
<td>0.7</td>
<td>560</td>
</tr>
<tr>
<td>May 16</td>
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</tr>
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<tr>
<td>May 22</td>
<td>1500</td>
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<td>675</td>
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</tbody>
</table>

*Date of first recovery: ANC ≥ 0.5×10⁹/L*

**Question 14: Was there evidence of initial hematopoietic recovery?**
Indicate whether or not there was evidence of initial ANC recovery following this HCT.
Check only one response:

- If Yes, ANC ≥ 500/mm³ (or ≥ 0.5 x 10⁹/L) achieved and sustained for 3 laboratory values, continue with question 15.
- If No, ANC ≥ 500/mm³ (or ≥ 0.5 x 10⁹/L) was not achieved, continue with question 116.
- Check Not applicable if the recipient’s ANC never dropped below 0.5 x 10⁹/L at any time post-HCT. This option is only applicable in the 100-day reporting period. Continue with question 16.
- Check Previously reported if this is the 6 month or annual follow-up, and the initial ANC recovery has already been reported. Continue with question 16.

When Not applicable is reported for 100-day reporting period, for all future reporting periods, select Previously reported.

Question 15: Date ANC ≥ 500/mm³ (first of 3 lab values)
Enter the first date of the three consecutive laboratory values obtained on different days where the ANC was ≥ 500/mm³ (or ≥ 0.5 × 10⁹/L). For an example of tracking ANC recovery, see the Tracking ANC Recovery example above.

For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

Question 16: Did late graft failure occur?
Late (or secondary) graft failure is defined when the recipient meets criteria for initial engraftment but subsequently develops loss of a previously functioning graft by development of at least two lines of cytopenia. Late graft failure is more often associated with allogeneic HCT than with autologous HCT. Some possible causes for late graft failure include graft rejection related to residual host immunity, persistent or progressive disease, low donor cell yield, medication side-effect, infection or GvHD.²

If the recipient meets the criteria of graft failure, check Yes.


Section Updates:

<table>
<thead>
<tr>
<th>Question Number</th>
<th>Date of Change</th>
<th>Add/Remove/Modify</th>
<th>Description</th>
<th>Reasoning (If applicable)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Q17 – 18: Initial Platelet Recovery

Initial Platelet Recovery
The Initial Platelet Recovery section can only be completed on the 100 day, 6 month, 1 year, and 2 year follow-up forms. These questions will be skipped for all subsequent reporting periods.

Optional for Non-U.S. Centers

The following questions refer to initial platelet recovery following the HCT for which this form is being completed. All dates should reflect no platelet transfusions administered for seven consecutive days. Report the date of the first of three consecutive laboratory values ≥ 20 × 10^9/L obtained on different days, as shown in the Reporting Platelet Recovery example below. Note that platelet recovery may take place well after the recipient has returned to the referring physician for care. It is essential that information and laboratory values be obtained from the referring physician.

Transfusions temporarily increase blood cell counts. When the data is later used for analysis, it is important to be able to distinguish between a recipient whose own body was creating the cells and a recipient who required transfusions to support the counts.

The following example illustrates the procedure to follow for reporting platelet recovery.

Reporting Platelet Recovery

<table>
<thead>
<tr>
<th>Transfusion</th>
<th>0</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
</tr>
</thead>
<tbody>
<tr>
<td>Platelet Count</td>
<td>10,000</td>
<td>35,000</td>
<td>30,000</td>
<td>25,000</td>
<td>10,000</td>
<td>15,000</td>
<td>19,000</td>
<td>23,000</td>
<td>25,000</td>
<td>40,000</td>
<td>50,000</td>
</tr>
</tbody>
</table>

Report 1/8/08 as date platelet count ≥ 20 × 10^9/L

Question 17: Was an initial platelet count ≥ 20 × 10^9/L achieved?
Indicate whether or not there was evidence of initial platelet recovery following this HCT.

Check only one response:

- If Yes, continue with question 18.
- If No, continue with question 19.
• Check **Not applicable**, if the recipient’s platelets never dropped below 20 x 10⁹/L at any time post-HCT and a platelet transfusion was never required. If the recipient’s platelet count drops below 20 x 10⁹/L and/or the recipient received a platelet transfusion even once, do not use this option. This option is only applicable in the 100-day reporting period. Continue with question 19.

• Check **Previously reported** if this is the 6 month or annual follow-up, and initial platelet recovery has already been reported on a previous form. Continue with question 19.

When **Not applicable** is reported for 100-day reporting period, for all future reporting periods, select **Previously reported**.

**Question 18: Date platelet ≥ 20 x 10⁹/L**
Enter the first date of three consecutive laboratory values obtained on different days where the platelet count was ≥ 20 × 10⁹/L. Ensure that no platelet transfusions were administered for seven days immediately preceding this date. Include day seven, as shown in the Reporting Platelet Recovery example above, when determining the recovery date.

**Reporting estimated dates:** If a recipient is not seen within a month after their last platelet transfusion, an estimated date may be reported. In this case, the date seven days after the last platelet transfusion may be reported (see example A below). However, if the recipient is seen within a month of the last platelet transfusion, an estimated date should not be reported.

If three laboratory values were not obtained on consecutive days, but a sequential rise of ≥ 20 x 10⁹/L is demonstrated, follow the examples below when determining an estimated date.

**Reporting Scenarios:**

**A.** The recipient is being seen in the outpatient clinic and receives a platelet transfusion on January 1. The platelet count is 22 × 10⁹/L on January 2, 24 × 10⁹/L on January 3, and 28 × 10⁹/L on January 4. The recipient does not come into the clinic for evaluation until one month later. The recipient has not received any more platelet transfusions and the platelet count is well above 20 × 10⁹/L. Report January 8 (day seven post-platelet transfusion) for the date of platelet recovery.

**B.** The recipient is being seen in the outpatient clinic and receives a platelet transfusion on January 1. The platelet count is ≥ 20 × 10⁹/L on January 2, January 3, and January 4. The recipient is then discharged back to their primary care physician. The transplant center receives a follow-up note from the primary care physician that states “recipient recovered their platelets in January of 2011.” Report
an estimated date of recovery using the guidelines available in General Instructions, General Guidelines for Completing Forms.

For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

Section Updates:

<table>
<thead>
<tr>
<th>Question Number</th>
<th>Date of Change</th>
<th>Add/Remove/Modify</th>
<th>Description</th>
<th>Reasoning (If applicable)</th>
</tr>
</thead>
</table>

**Q19 – 44: Graft versus Host Disease**

**Autologous Transplants**
If this was an autologous HCT, continue with the Liver Toxicity Prophylaxis section of the form. The graft-versus-host disease section should only be completed for allogeneic infusions.

Graft versus Host Disease (GVHD) is an immunological phenomenon resulting from the reaction of donor immune cells against major or minor histocompatibility antigens of the recipient. GVHD is primarily caused by donor-derived T-cells.

Factors influencing the severity of GVHD are related to three main categories: 1) donor or graft, 2) recipient, and 3) treatment. The most influential donor/graft factor is the degree of genetic disparity between the donor and the recipient (HLA match), but other risk factors include female donor to male recipient, donor parity, older donors, and T-cell dose. The occurrence of acute GVHD becomes a risk factor for the development of chronic GVHD. Recipient age and prior infections are also factors.

In the past, GVHD was classified as acute or chronic based on its time to diagnosis following transplant, and other clinical and histological (biopsy or post-mortem) features. Today, there has been increased recognition that acute and chronic GVHD are not dependent upon time since HCT, so determination of acute or chronic should rest on clinical and histologic features. However, organ staging, and overall grade should only be calculated from the clinical picture, not histology. Acute GVHD usually begins between 10 and 40 days after HCT but can appear earlier or later. The organs most commonly affected by acute GVHD are the skin, gut, or liver.
If acute GVHD is diagnosed prior to chronic GVHD, report the diagnosis information, maximum severity of any symptoms, and treatment administered up to the date of diagnosis of chronic GVHD in the acute GVHD section of the form (questions 19-36). Do not include any signs, symptoms, or treatment occurring on or after the onset of chronic GVHD when completing the acute GVHD section. Report any new or persistent acute GVHD symptoms occurring on or after the onset of chronic GVHD only in the chronic GVHD section. If chronic GVHD was diagnosed in a prior reporting period, report No for questions 19 and 21 in each subsequent reporting period. See reporting scenarios included in question 19.

Transaminitis
Previously, if the recipient only had transaminitis related to acute GVHD, this would have been reported as “stage 0” liver GVHD with an overall grade of “not applicable.” However, as of July 2021, isolated transaminitis should not be reported as acute GVHD. In this scenario, report No, acute GVHD did not develop or persist. If the recipient has transaminitis and other organs involved (i.e., skin rash), then report Yes, acute GVHD developed or persisted but do not report there was liver involvement.

Question 19: Did acute GVHD develop since the date of last report?
Questions 19 and 21 on the Post-TED (2450) Form are meant to capture whether the recipient had active symptoms of acute GVHD during the reporting period. If the recipient had active acute GVHD during the reporting period, either question 19 or question 21 must be answered Yes unless there has been a prior / concurrent diagnosis of chronic GVHD (see note above question 19). There will not be a situation where Yes is reported for both question 19 and question 21. If this question is answered as Yes and a diagnosis date has been reported, the question Did acute GVHD persist since the date of last report will be disabled in FormsNetSM. Centers should report Yes for this question to indicate the recipient developed acute GVHD in the following scenarios:

- Acute GVHD is diagnosed for the first time during the reporting period.
- An acute GVHD flare is diagnosed during the current reporting period and all of the following conditions are met:
  - The recipient’s prior acute GVHD symptoms did not persist from the prior reporting period into the beginning of the current reporting period.
  - The flare is diagnosed after at least 30 days without any active acute GVHD symptoms.
  - The recipient was not diagnosed with chronic GVHD on or before the date of the flare (see note above question 19).

If the recipient does have active acute GVHD during the reporting period, but does not match either of the scenarios above, the center will likely need to report No for question this question and Yes for the question Did acute GVHD persist since the date of last report. Question 21, Did acute GVHD persist since the date of last report, is intended to
capture acute GVHD which has continued from a prior reporting period. This includes any flares which do not meet the above conditions.

The intent of classifying GVHD episodes as newly developed or persistent is to avoid having centers re-report diagnosis information which has been captured on a prior form. Refer to the Acute GVHD Diagnosis Scenarios below to see examples of how to answer questions 19 and 21.

Report No for questions 19 and 21 if the recipient had no active acute GVHD symptoms during the reporting period OR all acute GVHD signs / symptoms during the reporting period occurred after a diagnosis of chronic GVHD (see note above question 19).

Indicate Unknown if there is no information about the recipient’s GVHD status for the reporting period. This option should be used sparingly and only when no judgment can be made about the presence or absence of GVHD in the reporting period.

**Acute GVHD Diagnosis Scenarios:**

A. A recipient receives a HCT on 1/1/2015 and develops acute GVHD which is clinically diagnosed on 2/1/2015. At least one of their symptoms, attributed to acute GVHD, persists beyond the 100 day date of contact which is 4/5/2015. Treatment continues and symptoms completely resolve on 5/1/2015. Immunosuppression is tapered until a flare of acute GVHD is diagnosed on 5/25/2015. Immunosuppression is given and symptoms quickly resolve with no active acute GVHD beginning 6/10/2015. The six month date of contact is 6/20/2015. Another flare of acute GVHD is clinically diagnosed on 8/15/2015.

**100 Day Post-TED Form:**

Question 19: Report “yes” to indicate a new clinical diagnosis of acute GVHD.
Question 20: Report the initial date of diagnosis (2/1/2015).
Question 21: Leave blank. This question will be skipped whenever a diagnosis date has been entered in question 20.
Questions 22 – 28: Answer these questions based on the assessments performed at the time of diagnosis (2/1/2015).

**Six Month Post-TED Form:**

Question 19: Report “no” to indicate GVHD persists from a previous report and, therefore, did not develop during the reporting period.
Question 20: Leave blank. This question will be skipped whenever question 19 is answered “no.”
Question 21: Report “yes” to indicate GVHD persists form a previous report.
Questions 22 – 28: Leave blank. Answering “yes” for question 19 prevents the
center from re-reporting diagnosis information already captured on the 100-day form.

One Year Post-TED Form:

Question 19: Report “yes” to indicate a flare of GVHD occurred at least 30 days after resolving during a prior reporting period.
Question 20: Report the diagnosis date of the flare occurring during the reporting period (8/15/2015).
Question 21: Leave blank. This question will be skipped whenever a diagnosis date has been entered in question 20.
Questions 22 – 28: Answer these questions based on the assessments performed at the time of diagnosis of the flare of acute GVHD (8/15/2015).

B. A recipient receives a HCT on 1/1/2015 and develops acute skin GVHD on 2/1/2015 and then chronic eye GVHD on 3/1/2015. Both acute and chronic symptoms resolve by the 100-day date of contact (4/5/2015). While tapering their immunosuppression, the recipient has a flare of their acute skin GVHD on 5/30/2015. Treatment continues and symptoms completely resolve by the six-month date of contact (6/20/2015).

100 Day Post-Infusion Data Form:

Question 19: Report “yes” to indicate a new clinical diagnosis of acute GVHD.
Question 20: Report the initial date of diagnosis (2/1/2015).
Question 21: Leave blank. This question will be skipped whenever a diagnosis date has been entered in question 20.
Questions 22 – 28: Answer these questions based on the assessments performed at the time of diagnosis (2/1/2015).
Questions 29 – 36: Answer these questions based on any symptoms and treatment documented from the onset of acute GVHD (2/1/2015) up to the diagnosis of chronic GVHD (3/1/2015). This instruction is provided in the note box above question 19.

Six Month Post-Infusion Data Form:

Question 19: Report “no” to indicate acute GVHD did not develop during the reporting period.
Question 20: Leave blank. This question will be skipped whenever question 19 is answered “no.”
Question 21: Report “no” to indicate acute GVHD did not persist from a previous report.

If chronic GVHD has been diagnosed in a prior reporting period, report “no” for questions 19 and 21. Any new or persistent acute GVHD symptoms occurring...
after the onset of chronic GVHD must be reported in the chronic GVHD section of the form. Do not include any signs, symptoms, or treatment occurring on or after the onset of chronic GVHD when completing the acute GVHD section. This instruction has been provided in the note above question 19.

Question 20: Date of acute GVHD diagnosis
Report the date of clinical diagnosis of acute GVHD. The clinical diagnosis date may not necessarily be the date the symptoms began (example: the recipient developed a rash one week prior to the physician clinically diagnosing acute skin GVHD). If the clinical diagnosis is documented, but the diagnosis date is unclear, obtain documentation from the primary physician confirming the clinical diagnosis date.

If the recipient developed more than one episode of acute GVHD in the same reporting period, report the date of onset of the first episode of acute GVHD.

For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

Question 21: Did acute GVHD persist since the date of last report?
This question will only be enabled in FormsNet3 if the center has reported No for the questions Did acute GVHD develop since the date of last report and, therefore, has not reported a date of diagnosis. If prompted to answer this question, report Yes if acute GVHD was diagnosed in a prior reporting period and any of the following conditions are met:

- The recipient’s acute GVHD symptoms have been active since diagnosis and continue to be active during the current reporting period (i.e., no period of resolution or quiescence since diagnosis).  
- The recipient’s acute GVHD symptoms had resolved before the first day of the current reporting period, but a flare occurred within 30 days of symptom resolution / quiescence.  
- The recipient was not diagnosed with chronic GVHD on or before the date of the flare (see note above question 19)

If Yes is reported for this question, go to question 29.

Report No for questions 19 and 21 if the recipient had no active acute GVHD symptoms during the reporting period OR all acute GVHD signs / symptoms during the reporting period occurred after a diagnosis of chronic GVHD (see note above question 19). If No is reported for this question, go to question 37.

Indicate Unknown if there is no information about the recipient’s GVHD status for the reporting period. This option should be used sparingly and only when no judgment can be made about the presence or absence of GVHD in the reporting period. If Unknown is reported, go to question 37.
Question 22: Overall grade of acute GVHD at diagnosis
Indicate the overall grade of acute GVHD at the time of diagnosis. For reporting purposes, “at diagnosis” is defined as the period between onset of signs / symptoms and the initiation of therapy to treat GVHD (topical or systemic). The acute GVHD grading scale is based on clinical evidence (physician observation), not histology. Pathology reports sometimes list a histologic grade of GVHD. Do not report the histologic grade. GVHD scoring and grading is based on clinical severity, not histologic severity. Biopsy of affected organs allows for more precise diagnosis as to the presence or absence of GVHD. However, overall grading remains clinical and is based on the criteria published by Przepiorka et al., Bone Marrow Transplant 1995; 15(6):825-8, see the GVHD Grading and Staging table below.

The CIBMTR will continue to collect overall grade of acute GVHD data based on the Przepiorka et al. criteria. New methods of grading acute GVHD, such as the MAGIC consortium criteria1, can be used internally at sites; however, all data reported to the CIBMTR should be consistent with the Przepiorka et al. criteria.

If acute GVHD was present, but the grade at diagnosis was not documented and it cannot be determined from the grading and staging table, report Not applicable.

Examples may include:
- Any other organ involvement without skin, liver, or gut symptoms attributable to GVHD
- Lower intestinal tract involvement where the stage cannot be determined in select scenarios (see lower intestinal tract involvement description below)

Upper GI GVHD
If the recipient only has upper GI GVHD during the reporting period, report this as overall grade II. This may differ from prior instructions regarding how to report upper GI GVHD.

GVHD Grading and Staging

<table>
<thead>
<tr>
<th>Stage</th>
<th>Skin</th>
<th>Liver</th>
<th>Gut</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Rash on &lt;25% of skin¹</td>
<td>Bilirubin 2-3 mg/dl²</td>
<td>Diarrhea &gt; 500 ml/day² or persistent nausea or vomiting³ Pediatric: 280-555 ml/m²/day or 10-19.9 mL/kg/day</td>
</tr>
</tbody>
</table>

2 Rash on 25-50% of skin | Bilirubin 3-6 mg/dl | Diarrhea >1000 ml/day  
*Pediatric:* 556-833 ml/m²/day or 20-30 mL/kg/day

3 Rash on >50% of skin | Bilirubin 6-15 mg/dl | Diarrhea >1500 ml/day  
*Pediatric:* >833 ml/m²/day or >30 mL/kg/day

4 Generalized erythroderma with bullous formation | Bilirubin >15 mg/dl | Severe abdominal pain, with or without ileus, and/or grossly bloody stool

### Grade

<table>
<thead>
<tr>
<th></th>
<th>Stage 1-2</th>
<th>None</th>
<th>None</th>
</tr>
</thead>
<tbody>
<tr>
<td>II</td>
<td>Stage 3</td>
<td>Stage 1</td>
<td>Stage 1</td>
</tr>
<tr>
<td>III</td>
<td>—</td>
<td>Stage 2-3</td>
<td>Stages 2-4</td>
</tr>
<tr>
<td>IV</td>
<td>Stage 4</td>
<td>Stage 4</td>
<td>—</td>
</tr>
</tbody>
</table>

1. Use “Rule of Nines” ([Percent Body Surfaces table](#)) or burn chart to determine extent of rash.

2. Range given as total bilirubin. Downgrade one stage if an additional cause of elevated bilirubin has been documented.

3. Volume of diarrhea applies to adults. For pediatric patients, the volume of diarrhea should be based on body surface area. Downgrade one stage if an additional cause of diarrhea has been documented.

4. Persistent nausea with or without histologic evidence of GVHD in the stomach or duodenum.

5. Criteria for grading given as minimum degree of organ involvement required to confer that grade.

6. Grade IV may also include lesser organ involvement with an extreme decrease in performance status.

### Questions 23 – 28: List the stage for each organ at diagnosis of acute GVHD.

**Skin:** Select the stage that reflects the body surface area involved with a maculopapular rash attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. See the [Percent Body Surfaces](#) table below to determine the percent of body surface area involved with a rash. Do not report ongoing rash not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.
Percent Body Surfaces

<table>
<thead>
<tr>
<th>Body Area</th>
<th>Percent</th>
<th>Total Percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Each Arm</td>
<td>9%</td>
<td>18%</td>
</tr>
<tr>
<td>Each Leg</td>
<td>18%</td>
<td>36%</td>
</tr>
<tr>
<td>Chest &amp; Abdomen</td>
<td>18%</td>
<td>18%</td>
</tr>
<tr>
<td>Back</td>
<td>18%</td>
<td>18%</td>
</tr>
<tr>
<td>Head</td>
<td>9%</td>
<td>9%</td>
</tr>
<tr>
<td>Pubis</td>
<td>1%</td>
<td>1%</td>
</tr>
</tbody>
</table>

**Lower GI GVHD and Stool Output Not Documented**

If diarrhea is attributed to acute GVHD during the reporting period, but the volume of stool output is not documented, leave the lower GI stage data field blank, override the FormsNet3 error as “not documented,” and specify the volume of stool output was not documented. In this case, report **Not applicable** for the overall grade unless stage 4 acute skin GVHD, stage 4 acute liver GVHD, or an extreme decrease in performance status or stage 2 or 3 acute liver GVHD was also documented at the time point being reported *(at diagnosis or maximum grade during the current reporting period).*

**Lower intestinal tract (use mL/day for adult recipients and mL/m²/day for pediatric recipients):** Select the stage that reflects the volume of diarrhea attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. Use mL/day for adult recipients and mL/m²/day for pediatric recipients. Input and output records may be useful in determining the volume of diarrhea. Do not report diarrhea ongoing but not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

**Upper intestinal tract:** Select the stage that reflects the presence of persistent nausea or vomiting attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. Do not report nausea or vomiting ongoing but not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

**Liver:** Select the stage that reflects the bilirubin level attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. Do not report hyperbilirubinemia ongoing but not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

**Other site(s) involved with acute GVHD:** Indicate whether acute GVHD affected an organ other than skin, upper GI, lower GI, or liver manifesting with hyperbilirubinemia. Report only other organ involvement at the time of acute GVHD diagnosis or flare in the reporting period. Do not report symptoms ongoing but not attributed to acute GVHD at the time of
acute GVHD diagnosis or flare. Specify the other organ system involvement in question 28.

**Question 29: Maximum overall grade of acute GVHD**
Indicate the overall maximum grade of acute GVHD since the date of the last report. Grading is based onclinical evidence (physician observation), not histology. Pathology reports sometimes list a histologic grade of GVHD. Do not report the histologic grade. GVHD scoring and grading is based on clinical severity, not histologic severity. Biopsy of affected organs allows for more precise diagnosis as to the presence or absence of GVHD. However, overall grading remains clinical and is based on the criteria published by Przepiorka et al., *Bone Marrow Transplant* 1995; 15(6):825-8; see the [GVHD Grading and Staging](#) table above.

If chronic GVHD was diagnosed during the reporting period, report the maximum severity of acute GVHD prior to the onset of chronic GVHD. See question 19 for further instructions. Acute GVHD grading scenario D below has been provided for further clarification.

Report the recipient’s maximum acute GVHD grade in the reporting period; this may differ from the grade at diagnosis or may be the same. If acute GVHD was present, but the maximum grade was not documented and it cannot be determined from the grading and staging table, report Not applicable.

Examples may include:
- Any other organ involvement without skin, liver, or gut symptoms attributable to GVHD
- Lower intestinal tract involvement where the stage cannot be determined in select scenarios (see lower intestinal tract involvement description above)

---

**Upper GI GVHD**
If the recipient only has upper GI GVHD during the reporting period, report this as overall grade II. This may differ from prior instructions regarding how to report upper GI GVHD.

**Acute GVHD Grading Scenarios:**

**A.** A recipient developed stage 2 skin involvement and elevated liver function tests (LFTs) attributed to acute GVHD; however, there was no total bilirubin manifestation. In this case, overall maximum grade I acute GVHD should be reported since the staging / grading can be determined using the [GVHD Grading and Staging](#) table above.

**B.** A recipient developed acute liver GVHD with elevated LFTs (i.e., transaminases) with no total bilirubin manifestation. The progress notes
indicate stage 1 (grade II overall) acute GVHD of the liver. In this case, this would not be reported as acute GVHD.

**C.** A recipient developed stage 2 skin involvement, which showed improvement in response to topical steroids. However, the recipient then developed hyperbilirubinemia attributed to stage 1 liver involvement; the skin involvement at that time was stage 1. In this case, grade II would be reported (assuming this was the extent of the recipient’s acute GVHD in the reporting period).

**D.** A recipient developed stage 2 skin involvement which resolved in response to topical steroids. Later in the reporting period, the recipient was diagnosed with mild chronic eye GVHD. Shortly thereafter, they were diagnosed with a stage 3 flare of acute skin GVHD. In this case, grade I would be reported. Do not consider any new or persistent acute GVHD symptoms occurring after the onset of chronic GVHD when completing the acute GVHD section of the form.

**E.** A recipient developed stage 1 skin involvement on 1/1/2019 which resolved in response to topical steroids and tacrolimus. Later in the reporting period, on 2/14/2019, they have a flare of the skin GVHD, this time at stage 2. In this case, grade I would be reported as the maximum overall grade of acute GVHD with the date of diagnosis of the more severe flare (2/14/2019). Additionally, the skin symptoms would be reported as stage 2 for the maximum skin stage.

**Question 30: Date maximum grade overall grade of acute GVHD**

Report the first date of maximum acute GVHD involvement, based on clinical grade. If the recipient had multiple instances in which their GVHD reached the same maximum grade, report the earliest date. However, if the same maximum overall grade was achieved, but the specific organ staging varied, report the date of the maximum organ staging which is consistent with the overall grade reported in question 25. Acute GVHD grading scenario E above has been provided for further clarification.

**Questions 31 – 36: List the stage for each organ at the time of maximum overall grade of acute GVHD.**

**Skin:** Select the stage that reflects the body surface area involved with a maculopapular rash attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. See the Percent Body Surfaces table below to determine the percent of body surface area involved with a rash. Do not report ongoing rash not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

### Percent Body Surfaces

<table>
<thead>
<tr>
<th>Body Area</th>
<th>Percent</th>
<th>Total Percentage</th>
</tr>
</thead>
</table>

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Lower intestinal tract (use mL/day for adult recipients and mL/m²/day for pediatric recipients): Select the stage that reflects the volume of diarrhea attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. Use mL/day for adult recipients and mL/m²/day for pediatric recipients. Input and output records may be useful in determining the volume of diarrhea. Do not report diarrhea ongoing but not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

Report an overall grade of IV if stage 4 acute skin GVHD, stage 4 acute liver GVHD, or an extreme decrease in performance status is documented at the time point being reported (see GVHD Staging and Grading Table). Report overall grade III if stage 2-3 liver involvement is documented at the time point being reported and there is no evidence of grade IV GVHD.

Upper intestinal tract: Select the stage that reflects the presence of persistent nausea or vomiting attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. Do not report nausea or vomiting ongoing but not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

Liver: Select the stage that reflects the bilirubin level attributed to acute GVHD at the time of acute GVHD diagnosis or flare in the reporting period. Do not report hyperbilirubinemia ongoing but not attributed to acute GVHD at the time of acute GVHD diagnosis or flare.

Other site(s) involved with acute GVHD: Indicate whether acute GVHD affected an organ other than skin, upper GI, lower GI, or liver manifesting with hyperbilirubinemia. Report only other organ involvement at the time of maximum overall grade of acute GVHD in the reporting period. Do not report symptoms ongoing but not attributed to acute GVHD at the time of maximum overall grade of acute GVHD. Specify the other organ system involvement in question 36.

Question 37: Did chronic GVHD develop since the date of last report? Indicate whether a new clinical diagnosis of chronic GVHD was documented during the reporting period. If chronic GVHD was diagnosed during the reporting period, report Yes.
If the recipient had a flare of chronic GVHD occurring after at least a 30-day period of symptom quiescence, report Yes. Report No if symptoms resolve or become quiescent prior to the date of last report and then flare within 30 days. This should be reported as persistent chronic GVHD which is captured in the question Did chronic GVHD persist since the date of last report.

Report No if chronic GVHD was not clinically diagnosed – initially or as a flare – in the reporting period; this includes instances where chronic GVHD persists from a prior reporting period.

Indicate Unknown if there is no information about the recipient’s GVHD status for the reporting period. This option should be used sparingly and only when no judgment can be made about the presence or absence of GVHD in the reporting period.

**Question 38: Date of chronic GVHD diagnosis**

Report the date of clinical diagnosis of chronic GVHD. The clinical diagnosis date may not necessarily be the date the symptoms began (example: the recipient developed shortness of breath one month prior to the clinical diagnosis of pulmonary chronic GVHD). If the clinical diagnosis is documented, but the diagnosis date is unclear, obtain documentation from the primary physician confirming the clinical diagnosis date.

If the recipient developed more than one episode of chronic GVHD in the same reporting period, report the date of onset of the first episode of chronic GVHD.

For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

**Question 39: Did chronic GVHD persist since the date of last report?**

Indicate whether chronic GVHD was clinically diagnosed during a previous reporting period and persisted, with active symptoms, into the present reporting period. Do not report quiescent or inactive chronic GVHD, or a prior history of GVHD. If Yes, questions concerning chronic GVHD at the time of diagnosis will be skipped. See question 33 for instructions on reporting a chronic GVHD flare.

If the recipient has no active symptoms during the reporting period, report No continue with question 43.

Indicate Unknown if there is no information about the recipient’s GVHD status for the reporting period. This option should be used sparingly and only when no judgment can be made about the presence or absence of GVHD in the reporting period.

**Question 40: Maximum grade of Chronic GVHD (according to best clinical judgement)**
Report the maximum chronic GVHD involvement, based on clinical grade, since the
date of the last report. The intent of this question is to capture the maximum grade
based on the best clinical judgment. If the maximum clinical grade is not documented,
request documentation from the recipient’s primary care provider. Guidelines on how to
report the maximum grade of chronic GVHD are outlined below:

- **Mild**: Signs and symptoms of chronic GVHD do not interfere substantially with
  function and do not progress once appropriately treated with local therapy or
  standard systemic therapy (e.g. corticosteroids and/or cyclosporine or FK 506)

- **Moderate**: Signs and symptoms of chronic GVHD interfere somewhat with
  function despite appropriate therapy or are progressive through first line systemic
  therapy (e.g. corticosteroids and/or cyclosporine or FK 506)

- **Severe**: Signs and symptoms of chronic GVHD limit function substantially
despite appropriate therapy or are progressive through second line therapy.

### Organ Scoring of Chronic GVHD

<table>
<thead>
<tr>
<th>Organ</th>
<th>Score 0</th>
<th>Score 1</th>
<th>Score 2</th>
<th>Score 3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Skin % BSA</td>
<td>No BSA involved</td>
<td>1-18% BSA</td>
<td>19-50% BSA</td>
<td>&gt;50% BSA</td>
</tr>
<tr>
<td>Skin Features</td>
<td>No sclerotic features</td>
<td>N/A</td>
<td>Superficial sclerotic features, but not “hidebound”</td>
<td>Deep sclerotic features; “hidebound;” impaired mobility; ulceration</td>
</tr>
<tr>
<td>Mouth</td>
<td>No symptoms</td>
<td>Mild symptoms with disease signs but not limiting oral intake significantly</td>
<td>Moderate symptoms with disease signs with partial limitation of oral intake</td>
<td>Severe symptoms with disease signs with major limitation of oral intake</td>
</tr>
<tr>
<td>Eyes</td>
<td>No symptoms</td>
<td>Mild dry eye symptoms not affecting ADL (requirement of lubricant drops ≤ 3x/day)</td>
<td>Moderate dry eye symptoms partially affecting ADL(requiring lubricant drops &gt; 3x/day or punctal plugs) WITHOUT new vision impairment due to keratoconjunctivitis sicca (KCS)</td>
<td>Severe dry eye symptoms significantly affecting ADL(special eyewear to relieve pain) OR unable to work because of ocular symptoms OR loss of vision due to keratoconjunctivitis sicca (KCS)</td>
</tr>
<tr>
<td>GI Tract</td>
<td>No symptoms</td>
<td>Symptoms without significant weight loss (&lt; 5%)</td>
<td>Symptoms associated with mild to moderate weight loss (5-15%) within 3 months OR moderate diarrhea without</td>
<td>Symptoms associated with significant weight loss (&gt; 15%) within 3 months, requires nutritional supplement for most calorie</td>
</tr>
<tr>
<td>Liver</td>
<td>Normal total bilirubin and ALT or AP &lt; 3 x ULN</td>
<td>Normal total bilirubin with ALT ≥ 3 to 5 x ULN or AP ≥ 3 x ULN</td>
<td>Elevated total bilirubin but ≤ 3 mg/dL or ALT &gt; 5 x ULN</td>
<td>Elevated total bilirubin &gt; 3 mg/dL</td>
</tr>
<tr>
<td>-------</td>
<td>-----------------------------------------------</td>
<td>---------------------------------------------------------------</td>
<td>-----------------------------------------------------</td>
<td>----------------------------------</td>
</tr>
<tr>
<td>Lungs Symptom Score:</td>
<td>No symptoms</td>
<td>Mild symptoms (SOB after climbing one flight of steps)</td>
<td>Moderate symptoms (SOB after walking on flat ground)</td>
<td>Severe symptoms (SOB at rests; requires O2)</td>
</tr>
<tr>
<td>Lung Score:</td>
<td>FEV1 ≥ 80%</td>
<td>FEV1 60-79%</td>
<td>FEV1 40-59%</td>
<td>FEV1 ≤ 39%</td>
</tr>
<tr>
<td>Joints and Fascia</td>
<td>No symptoms</td>
<td>Mild tightness of arms or legs, normal or mild decreased range of motion AND not affecting ADL</td>
<td>Tightness of arms or legs OR joint contractures, erythema thought to be due to fasciitis, moderate decrease of range of motion AND mild to moderate limitation of ADL</td>
<td>Contractures WITH significant decrease of range of motion AND significant limitation of ADL (unable to tie shoes, button shirts, dress self, etc.)</td>
</tr>
<tr>
<td>Genital Tract²</td>
<td>No signs</td>
<td>Mild signs and females with or without discomfort on exam</td>
<td>Moderate signs and may have signs of discomfort on exam</td>
<td>Severe signs with or without symptoms</td>
</tr>
<tr>
<td>Other Features³</td>
<td>No GVHD</td>
<td>Mild</td>
<td>Moderate</td>
<td>Severe</td>
</tr>
</tbody>
</table>

NIH Consensus Criteria, 2014

1. Features to be scored by BSA: Maculopapular rash, lichen planus-like features, sclerotic features, papulosquamous lesions or ichthyosis, keratosis pilaris-like GVHD.

2. Scoring is based on severity of the signs instead of symptoms, based on limited available data and the opinions of experts. Female or male genital GVHD is not scored if a practitioner is unable to examine the patient.

3. May include ascites, pericardial effusion, pleural effusion(s), nephrotic syndrome, myasthenia gravis, peripheral neuropathy, polymyositis, weight loss without GI symptoms, eosinophilia > 500/μL, platelets < 100,000/μL, others

Question 41: Specify if chronic GVHD was limited or extensive
The grading system for chronic GVHD is divided into two categories: limited and extensive. Definitions are based on Sullivan KM, Blood 1981; 57:267.
Report **Limited** if chronic GVHD includes only localized skin involvement and/or liver dysfunction. Report **Extensive** if **any** of the following symptoms are attributed to chronic GVHD:

- Generalized skin involvement and/or liver dysfunction
- Liver histology showing chronic aggressive hepatitis, bridging necrosis, or cirrhosis
- Involvement of the eye: Schirmer’s test with < 5 mm wetting, or
- Involvement of minor salivary glands or oral mucosa, or
- Involvement of any other target organ

**Question 42: Date of maximum grade of chronic GVHD**
Report the date of maximum chronic GVHD involvement, based on clinical grade, during the current reporting period. If the recipient had multiple instances in which their GVHD reached the same maximum grade, report the earliest date.

For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

**Question 43: Is the recipient still taking systemic steroids?** (Do not report steroids for adrenal insufficiency, or steroid dose ≤10 mg/day for adults, <0.1 mg/kg/day for children)

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**Steroids and Non-Steroid Immunosuppression for GVHD**
GVHD treatment questions will only be completed if the center has reported **Yes** acute and/or chronic GVHD develop or persisted since the date of last report. If **No** has been reported, then the GVHD treatment questions will be left blank.
Indicate whether the recipient is still taking systemic steroids to treat or prevent GVHD on the date of contact. Refer to the guidelines included in the question text if the recipient is taking low dose steroids or steroids for adrenal insufficiency.

Indicate **Not applicable** in any of the following scenarios:

- The recipient has never received systemic steroids (> 10 mg / day for adults or ≥ 0.1 mg / kg / day for children) to treat or prevent GVHD.
- This form is being completed for a subsequent HCT and the recipient has never received systemic steroids (> 10 mg / day for adults or ≥ 0.1 mg / kg / day for children) to treat or prevent GVHD since the start of the preparative regimen for the most recent infusion (or since the date of the most recent infusion if no preparative regimen is given).
- The recipient stopped taking systemic steroids (> 10 mg / day for adults or ≥ 0.1 mg / kg / day for children) to treat or prevent GVHD in a previous reporting period and did not restart systemic steroids (> 10 mg / day for adults or ≥ 0.1 mg / kg / day for children) during the current reporting period.

Indicate **Unknown** if there is no information to determine if the recipient is still taking systemic steroids. This option should be used sparingly and only when no judgment can be made about the recipient still receiving treatment for GVHD on the date of contact.

If the recipient has died prior to the discontinuation of systemic steroids used to treat or prevent acute and / or chronic GVHD, select **Yes**.

**Question 44: Is the recipient still taking (non-steroid) immunosuppressive agents (including PUVA) for GVHD?**

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**Corticosteroids**

Corticosteroids are captured differently depending on whether they are used topically or systemically. Use the following guidelines when determining how to report corticosteroids used to treat acute GVHD:

- **Topical Creams for Skin:** Do not report topical ointments or creams used to treat skin GVHD including corticosteroid creams such as Triamcinolone or Hydrocortisone.
- **Other Topical Treatments:** Certain corticosteroid treatments are inhaled or ingested but are not absorbed and are therefore considered topical. Examples include beclomethasone and budesonide. Do not consider these medications when answering the question regarding systemic steroids.
- **Systemic Treatments:** Systemic administration of corticosteroids, including use of prednisone and dexamethasone, should be reported in the question regarding systemic steroids.
GVHD treatment questions will only be completed if the center has reported Yes acute and/or chronic GVHD develop or persisted since the date of last report. If No has been reported, then the GVHD treatment questions will be left blank.

Indicate whether the recipient is still taking non-steroidal immunosuppressive agents (including PUVA) to treat or prevent acute and/or chronic GVHD on the date of contact. Descriptions of many immunosuppressive agents are included below.

If the recipient did not receive non-steroidal immunosuppressive agents to treat or prevent acute and/or chronic GVHD during the reporting period, report Not applicable.

Indicate Not applicable in any of the following scenarios:

- The recipient has never received non-steroidal immunosuppressive agents (including PUVA) to treat or prevent GVHD.
- This form is being completed for a subsequent HCT and the recipient has never received non-steroidal immunosuppressive agents (including PUVA) to treat or prevent GVHD since the start of the preparative regimen for the most recent infusion (or since the date of the most recent infusion if no preparative regimen was given).
- The recipient stopped taking non-steroidal immunosuppressive agents (including PUVA) to treat or prevent GVHD in a previous reporting period and did not restart non-steroidal immunosuppressive agents (including PUVA) during the current reporting period.

Indicate Unknown if there is no information to determine if the recipient is still taking non-steroidal immunosuppressive agents. This option should be used sparingly and only when no judgment can be made about the recipient still receiving treatment for GVHD in the reporting period.

Immunosuppressive Agents:

**Aldesleukin (Proleukin):** Increases production of several white blood cells including regulatory T-cells. This drug is also known as interleukin-2.

**ALG (Anti-Lymphocyte Globulin), ALS (Anti-Lymphocyte Serum), ATG (Anti-Thymocyte Globulin) ATS (Anti-Thymocyte Serum):** Serum or gamma globulin preparations containing polyclonal immunoglobulins directed against lymphocytes. These drugs are usually prepared from animals immunized against human lymphocytes. Also report the animal source. If “other” is selected, specify the source.

**Azathioprine (Imuran):** Azathioprine inhibits purine synthesis. Usually it is used at low doses in combination with other treatments.

**Bortezomib (Velcade):** A proteasome inhibitor.
**Cyclosporine (CSA, Neoral, Sandimmune):** Calcineurin inhibitor which decreases cytokine production by T-cells. Usually given for ≥ 3 months.

**Cyclophosphamide (Cytoxan):** Given in high doses near the date of infusion as single agent prophylaxis.

**Extra-corporeal photopheresis (ECP):** The recipient’s blood is removed from the body, exposes to psoralen and ultraviolet light, and re-infused.

**FK 506 (Tacrolimus, Prograf):** Inhibits the production of interleukin-2 by T-cells.

**Hydroxychloroquine (Plaquenil):** Hydroxychloroquine inhibits transcription of DNA to RNA and is commonly used as an anti-malarial drug.

**Interleukin Inhibitor:** Interleukin inhibitors suppress production of white blood cells and are grouped according to their target. Examples of IL-2 inhibitors include daclizumab (Zynbryta) and basiliximab (Simulect). Examples of IL-6 inhibitors include tocilizumab (Actemra) and siltuximab (Sylvant).

**In vivo monoclonal antibody:** Antibody preparations that are infused in the recipient following HSCT. Specify the antibody used as: anti CD25 (Zenapax, Daclizumab, Anti-TAC), alemtuzumab (Campath), entanercept (Enbrel), infliximab (Remicade), and / or rituximab (Rituxan).

**In vivo immunotoxin:** Antibody preparations linked to a toxin that is infused in the recipient following HCT. Specify the immunotoxin.

**Janus Kinase 2 Inhibitors:** Suppress function of T-effector cells. Examples: ruxolitinib (Jakafi, Jakavi) and tofacitinib (Xeljanz, Jakvinus).

**Methotrexate (MTX) (Amethopterin):** Inhibits the metabolism of folic acid. It is most often used with cyclosporine and is usually for a short duration of time.

**Mycophenolate mofetil (MMF) (CellCept, Myfortic):** Inhibits the de novo pathway used for lymphocyte proliferation and activation.

**Pentostatin (Nipent):** Inhibits adenosine deaminase, which blocks DNA (and some RNA) synthesis.

**Sirolimus (Rapamycin, Rapamune):** Inhibits the response to interleukin-2, blocking the activation of T-cells.

**Tyrosine Kinase Inhibitor (TKI):** Suppress function of tyrosine kinases thereby downregulating the function of many other cellular proteins / processes including fibrosis and inflammation. Examples: imatinib (Gleevec, Glivec), nilotinib (Tasigna), and dasatinib (Sprycel).

**UV Therapy:** UVA or UVB radiation administered to affected areas of the skin in order to suppress proliferation of cells responsible for GVHD.

**PUVA (Psoralen and UVA):** Psoralen is applied or taken orally to sensitize the skin, and then the skin is exposed to UVA radiation.
UVB: Broadband- or Narrowband-UVB radiation is applied to the affected areas of the skin.

Section Updates:

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<thead>
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<th>Description</th>
<th>Reasoning (If applicable)</th>
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**Q45 – 47: Liver Toxicity Prophylaxis**

Liver Toxicity Prophylaxis section can only be completed on the 100 day and 6 month follow-up forms. These questions will be skipped for all subsequent reporting periods.

**Question 45: Was specific therapy used to prevent liver toxicity?**

Liver toxicities in transplant patients may be related to drugs / treatments, infection, GVHD, iron overload, cirrhosis, or sinusoidal obstructive syndrome (SOS) / veno-occlusive disease (VOD). Agents such as ursodiol may be given as prophylaxis against one or more of these transplant-related liver injuries. Agents given to prevent liver toxicity will generally be started prior to or during the conditioning regimen and may be continued well after transplant.

Indicate whether the recipient received any therapy intended to prevent liver toxicity during the reporting period, including therapy given during the conditioning regimen. Report only agents given to prevent liver toxicities, not those given to treat a diagnosed liver injury or toxicity. If liver toxicity prophylaxis was given, report **Yes**. If liver toxicity prophylaxis was not given during the reporting period, report **No** and continue with question 44.

**Questions 46 – 47: Specify therapy (check all that apply)**

Select the agent(s) given during the reporting period to prevent liver toxicity, including therapy given during the conditioning regimen. Only report agents given to prevent liver toxicities, not those given to treat a diagnosed liver injury or toxicity. If **Other** therapy is selected, specify agent(s).

Section Updates:

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Q48 – 49: Veno-occlusive disease (VOD) / Sinusoidal obstruction syndrome (SOS)

Veno-occlusive disease (VOD) / Sinusoidal obstruction syndrome (SOS) occurs following injury to the hepatic venous endothelium, resulting in hepatic venous outflow obstruction due to occlusion of the hepatic venules and sinusoids. This typically results in a distinctive triad of clinical signs including hepatomegaly with right upper quadrant tenderness, third space fluid retention (e.g., ascites), and jaundice with a cholestatic picture. For more information on VOD / SOS including diagnostic criteria, refer to the VOD / SOS section of the Forms Instructions Manual.

Question 48 – 49: Did veno-occlusive disease (VOD) / sinusoidal obstruction syndrome (SOS) develop since the date of last report?
Indicate whether VOD / SOS was diagnosed during the reporting period. If Yes, report the date of diagnosis. If VOD / SOS persisted from the prior reporting period, indicate No and go to question 50.

For more information regarding reporting partial or unknown dates, see General Instructions, General Guidelines for Completing Forms.

Section Updates:

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<th>Add/Remove/Modify</th>
<th>Description</th>
<th>Reasoning (If applicable)</th>
</tr>
</thead>
</table>
| Q50 – 56: Infection

Diagnosis of COVID-19 after the start of the preparative regimen: Any COVID-19 infections diagnosed after the start of the preparative regimen should be reported in questions 50 – 51 on the Post-TED (2450) form. An associated Respiratory Virus Post-Infusion Data (2149) form will be generated.

Reporting Multiple COVID-19 Infections
If a recipient was diagnosed with COVID-19 multiple times within the current reporting period, complete questions 46 – 47 for each diagnosis.
Questions 50 – 51: Did the recipient develop COVID-19 (SARS-CoV-2) since the date of last report?

SARS-CoV-2 is a novel virus belonging to the coronavirus (CoV) family that emerged in December 2019. The disease caused by this new CoV is known as COVID-19 (coronavirus disease 2019). The new virus is highly contagious and was officially declared a pandemic in March 2020. Transmission is believed to be from person to person through respiratory droplets from coughing and sneezing. Testing for COVID-19 is generally performed on specimens collected from a nasal swab or sputum sample.

As a result of the global COVID-19 pandemic, the U.S. Food and Drug Administration granted Sherlock Biosciences an emergency use of authorization (EUA) for its COVID-19 diagnostic assay, CRISPR. Although still in its infancy in real-life application, positive results by this method should be reported, even if tandem testing by other method(s) (i.e., PCR) indicate a negative result. If the CRISPR results are unclear, seek physician clarification.

Indicate whether or not the recipient has ever had a known COVID-19 (SARS-CoV-2) infection, based on a positive test result, at any during the current reporting period.

If the recipient has had a documented COVID-19 (SARS-CoV-2) infection, report Yes and then specify the first date of the pathological diagnosis in question 51.

If the recipient has not had a documented COVID-19 (SARS-CoV-2) infection, report No and continue with question 52.

Question 52: Was a vaccine for COVID-19 (SARS-CoV-2) received?
Indicate if the recipient received a vaccine for COVID-19 (one dose without a planned second dose, first dose with planned second dose, second dose, third dose, and / or booster dose) within the current reporting period.

If the recipient did not receive a vaccine for COVID-19 or it is not known if the recipient received a vaccine, select No or Unknown, respectively, and continue with question 57.

COVID-19 Vaccine Doses
FormsNet3SM application: Complete questions 53 – 56 to report all COVID-19 vaccine doses received in the current reporting period by adding an additional instance in the FormsNet3SM application. A separate instance should be added for each dose.

Paper form submission: Copy questions 53 – 56 and complete report all COVID-19 vaccine doses received in the current reporting period. A separate instance should be completed for each dose.

Questions 53 – 54: Specify vaccine brand
For the reported dose, specify the vaccine brand the recipient received. If the vaccine brand is not listed, select Other type and specify in question 54.
Questions 55 – 56: Select dose(s) received
For the reported dose, specify the vaccine dose the recipient in the current reporting period and specify the date when the dose was received. If the exact date is not known, use the process described in the General Instructions, Guidelines for Completing Forms and select Date estimated.

Section Updates:

<table>
<thead>
<tr>
<th>Question Number</th>
<th>Date of Change</th>
<th>Add/Remove/Modify</th>
<th>Description</th>
<th>Reasoning (If applicable)</th>
</tr>
</thead>
<tbody>
<tr>
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</tr>
</tbody>
</table>

Q57 – 64: New Malignancy, Lymphoproliferative or Myeloproliferative Disorder

Question 57: Did a new malignancy, myeloproliferative, or lymphoproliferative disease / disorder occur that is different from the disease / disorder for which the HCT or cellular therapy was performed? (include clonal cytogenetic abnormalities, and post-transplant lymphoproliferative disorders)
Indicate whether a new or secondary malignancy, lymphoproliferative disorder, or myeloproliferative disorder has developed. Do not report recurrence, progression, or transformation of the recipient’s primary disease (disease for which the transplant was performed), or relapse of a prior malignancy.

New malignancies, lymphoproliferative disorders, or myeloproliferative disorders include but are not limited to:

- Skin cancers (basal, squamous, melanoma)
- New leukemia
- New myelodysplasia
- Solid tumors
- PTLD (post-transplant lymphoproliferative disorder) report as lymphoma or lymphoproliferative disease

The following should not be reported as new malignancy:

- Recurrence of primary disease (report as relapse or disease progression)
- Relapse of malignancy from recipient’s pre-HCT medical history
- Breast cancer found in other (i.e., opposite) breast (report as relapse)
• Post-HCT cytogenetic abnormalities associated with the pre-HCT diagnosis (report as relapse)
• Transformation of MDS to AML post-HCT (report as disease progression)

**Recurrent Skin Cancers**
For most malignancies, do not report recurrence, progression or transformation of the recipient’s primary disease (disease for which the transplant was performed) or relapse of a prior malignancy in the “New Malignancy” section.

For example, a recipient had a basal cell skin cancer diagnosed on the neck four months post-HCT and six months later had another basal cell located on the nose. The lesion on the nose is not considered a metastasis from the neck, but a new discrete lesion. These discrete episodes should be reported as Basal cell skin malignancy questions on the Post-TED forms (revision 6, questions 58 – 64).

If a new malignancy, lymphoproliferative disorder, or myeloproliferative disorder was diagnosed during the reporting period, report Yes. If No, continue with question 65.

**Reporting Multiple New Malignancy, Myeloproliferative Disease / Disorders:**
*FormsNet3SM application*: Complete questions 58 – 64 for each new malignancy diagnosed since the date of last report by adding an additional instance in the *FormsNet3SM application*.
*Paper form submission*: Copy questions 58 – 64 for each new malignancy diagnosed since the date of last report.

**Questions 58 – 59: Specify new malignancy**
If the new malignancy or disorder does not fit into one of the categories specified, indicate Other new malignancy and specify.

**Question 60: Is the tumor EBV positive?**
If the disorder is lymphoma or lymphoproliferative disease, indicate if the tumor is EBV positive. This question only applies if Hodgkin lymphoma, Non-Hodgkin lymphoma, or Post-transplant lymphoproliferative disorder (PTLD) is reported as the new malignancy.

**Question 61: Date of diagnosis**
Report the date of first pathological diagnosis (e.g., biopsy) of the new malignancy. Enter the date the sample was collected for examination. If the diagnosis was determined at an outside center, and no documentation of a pathological or laboratory assessment is available, the dictated date of diagnosis within a physician note may be reported. Do not report the date symptoms first appeared.
For malignancies or disorders without pathologic diagnosis, report the date of clinical diagnosis or date of specimen collection for laboratory assessment confirming diagnosis.

If exact date of diagnosis is not known, refer to General Instructions, General Guidelines for Completing Forms, for information about reporting partial or unknown dates.

**Question 62: Was documentation submitted to the CIBMTR? (e.g. pathology / autopsy report or other documentation)**
Indicate whether documentation of the new malignancy, lymphoproliferative disorder, or myeloproliferative disorder was submitted to CIBMTR (e.g., pathology report, autopsy report).

The submission of a pathology report or other supportive documentation for each reported new malignancy is strongly recommended.

For further instructions on how to attach documents in FormsNet3SM, refer to the training guide.

**Question 63 – 64: Was the new malignancy donor / cell product derived?**
Indicate whether the new malignancy originated from the donor / cell product. If Yes, indicate whether documentation was submitted to CIBMTR (e.g., cell origin evaluation (VNTR, cytogenetics, FISH)).

For further instructions on how to attach documents in FormsNet3SM, refer to the training guide.

**Q65 – 84: Chimerism Studies (Cord Blood Units, Beta Thalassemia, and Sickle Cell Disease Only)**

**Chimerism Studies**
This section relates to chimerism studies from allogeneic infusions using cord blood units, or for allogeneic infusion recipients whose primary disease is beta thalassemia or sickle cell disease only. If this was an autologous infusion, an allogeneic infusion using a bone marrow or PBSC product, and / or allogeneic infusion recipient whose primary disease for transplant was not beta thalassemia or sickle cell disease, continue to the disease assessment section.

**Chimerism Studies**
Chimerism study questions can only be completed on the 100-day, 6 month, 1 year, and 2 year follow-up forms. These questions will be skipped for all subsequent reporting periods.
Chimerism studies are performed to determine the percent of blood or marrow cells post-transplant that are produced from donor hematopoietic stem cells and the percent that are produced from host (recipient) hematopoietic stem cells. Different types of blood cells and a variety of laboratory tests can be used to determine if a chimera (presence of both donor- and host-derived cells) exists. If cytogenetic testing was performed to look for disease markers, and the donor and recipient are different sexes, the test may also be used to determine if a chimera exists. If the donor and recipient are of the same sex, cytogenetic testing using the common staining technique, known as giemsa banding (G-banding), cannot be used to determine if there is a chimera. However, quinicrine banding (Q-banding) can be used to identify if the cells are of donor origin or not in a same-sex transplant, as this staining technique highlights inherited chromosome polymorphisms on certain human chromosomes including 3, 4, 13, 15, 21, 22, and Y. This is not a commonly used staining technique and is only helpful when the polymorphism is documented pre-HCT.

Chimerism Studies
If chimerism studies were attempted, but no evaluable results were obtained, do not report the test.

When a multi-donor chimera exists and includes a donor (or donors) from a previous infusion, report as a multi-donor chimera though there may only be one donor for the current transplant.

Question 65 – 66: Were chimerism studies performed since the date of last report?
Indicate whether chimerism studies were performed within the reporting period. If Yes, indicate whether documentation was submitted to CIBMTR (e.g., chimerism laboratory reports).

If chimerism studies were not performed within the reporting period, select No and continue with question 85.

Question 67: Were chimerism studies assessed for more than one donor / multiple donors?
Indicate whether this HCT included product(s) from multiple donors. When a multi-donor chimerism exists and includes a donor or donors from a previous HCT, report as a multi-donor chimerism even though there may only be one donor for the current transplant.

Reporting Multiple Chimerism Studies:
**FormsNet3℠ application:** Complete questions 68 – 84 for each chimerism study by adding an additional instance in the FormsNet3℠ application.
**Paper form submission:** Copy questions 68 – 84 for each chimerism study since the date of last report.
Question 68 – 84: Provide date(s), method(s) and other information for all chimerism studies performed since the date of last report

When reporting chimerism studies for multiple donors, there should be one instance for each donor for each chimerism test result.

Transplant centers may perform frequent chimerism studies. If there is a need to reduce the number of chimerism study results reported due to volume, ensure that the following are reported at a minimum:

- Studies performed on or at approximately Day+28
- Most recent studies performed prior to the date of contact, particularly for Day+100
- Most recent studies performed prior to and after an intervention (such as a donor cellular infusion)
- The first result to show complete / 100% donor chimerism

### Chimerism - Single Donor

<table>
<thead>
<tr>
<th>Data Field</th>
<th>Description</th>
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<tbody>
<tr>
<td>68. NMDP donor ID</td>
<td>If the donor or one of the donors was an NMDP PBSC or marrow donor, enter the 9 digit NMDP donor ID.</td>
</tr>
<tr>
<td>69. NMDP cord blood unit ID</td>
<td>If the donor or one of the donors was an NMDP cord blood unit, enter the 9-digit NMDP cord blood unit ID.</td>
</tr>
<tr>
<td>70. Registry donor ID</td>
<td>If the donor was a non-NMDP donor, enter the registry donor ID.</td>
</tr>
<tr>
<td>71. Non-NMDP cord blood unit ID</td>
<td>If the donor or one of the donors was a non-NMDP cord blood unit, enter the non-NMDP registry donor ID.</td>
</tr>
<tr>
<td>72. Global Registration Identifiers for Donors (GRID)</td>
<td>The GRID standard (ICCBBA ST-015) is a 19-character donor identifier used to ensure that each donor ID is globally unique. For more information about the GRID, see the Pre-TED (2400) Forms Instruction Manual.</td>
</tr>
<tr>
<td>73. Donor date of birth or age</td>
<td>If the donor was related or the cord blood unit was related or supplied by a non-NMDP registry, provide the date of birth, if known; if date of birth is not known, provide the donor’s age at donation.</td>
</tr>
<tr>
<td>74. Sex</td>
<td>If the donor was related or the cord blood unit was related or supplied by a non-NMDP registry, provide the biological sex.</td>
</tr>
<tr>
<td>75. Date sample collected</td>
<td>Enter the date the sample was collected for the chimerism test.</td>
</tr>
<tr>
<td>76 – 77. Method</td>
<td>Report the test method used for the reported chimerism study. Cytogenetic testing methods include karyotyping and fluorescent in situ hybridization (FISH). Cytogenetic methods are only valid for sex mismatched transplants with the exception of quinicrine banding. VNTR / STR is one of the most common</td>
</tr>
</tbody>
</table>
molecular methods for assessing chimerism. See the Chimerism Methods table below for additional details on chimerism testing methods.

78. Cell source

Report whether the specimen taken for chimerism testing was from a Bone marrow or Peripheral blood source.

79 – 80. Cell type

Indicate the cell type tested. If the specimen was not sorted for a specific cell line, indicate Unsorted / whole. See the Chimerism Cell Types table below for additional details on cell markers unique to certain cell lines.

81. Total cells examined

Cytogenetic testing methods include karyotyping and fluorescent in situ hybridization (FISH), each of which examines a specific and relatively low number of cells – generally 15 to 200, depending on specimen and test method. If a cytogenetic method was used, enter the total number of cells that were examined. If a non-cytogenetic test was used, leave these boxes blank.

82. Number of donor cells

Cytogenetic methods, karyotyping and FISH, examine a specific and relatively low number of cells – generally 15 to 200, depending on specimen and test method. If a cytogenetic method was used, enter the total number of cells that were examined and found to be of donor origin. If a non-cytogenetic test was used, leave these boxes blank.

83. Were donor cells detected?

Molecular testing methods include RFLP and VNTR / STR. If a molecular method was used, indicate whether donor cells were detected. Report Yes, if the testing identified any percentage of cells as being of donor origin.

84. Percent donor cells

Molecular testing methods include VNTR / STR, RFLP, and AFLP. Report the percentage of donor cells identified by molecular testing. If the test result did not detect any recipient cell population within the sensitivity of the assay, report 100% donor cells. If the test detected recipient cells, but indicated donor cells "> n%,” report “n + 1” percent donor cells. If the test detected donor cells but indicated donor cells “< n%,” report “n – 1” percent donor cells.

### Chimerism Methods

<table>
<thead>
<tr>
<th>Method</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Karyotyping for XX / XY</td>
<td>Cells are grown in culture, stained, and examined under a microscope to identify the number of cells matching the sex of the donor. This method is only valid when donor and recipient are sex mismatched.</td>
</tr>
<tr>
<td>Fluorescent in situ hybridization (FISH) for XX / XY</td>
<td>Cells are exposed to fluorescent DNA probes which attach to X and Y chromosomes. A microscope is used to identify the number of cells matching the sex of the donor. This method is</td>
</tr>
</tbody>
</table>
only valid when donor and recipient are sex mismatched. Do not report FISH testing for disease-specific abnormalities in the chimerism section of the Post-TED.

| Restricted fragment length polymorphisms (RFLP) | A restriction fragment is a portion of DNA which has been cut out by an enzyme. RFLP testing begins by isolating DNA from the sample. Enzymes are used to cut the DNA at specific loci resulting in many unique restriction fragments. The fragments are separated according to size by electrophoresis. The unique pattern of separation is used to identify the percent donor DNA present in the sample. |
| Variable number tandem repeat (VNTR), micro- or minisatellite | VNTR refers to a portion of DNA containing a repeating sequence of base pairs (micro- or minisatellite). The number of times a micro- or minisatellite repeats within specific loci can differ between individuals. These differences are used to distinguish donor DNA from recipient DNA. VNTR testing involves obtaining samples from the recipient and donor prior to transplant. Specific loci are compared to determine which loci contain VNTRs unique to the donor. After transplant, DNA is isolated from recipient samples. Donor-specific VNTRs are amplified by PCR techniques. The sample is then analyzed to determine the percent donor DNA present. |
| Small tandem repeat (STR), micro- or minisatellite | STR also refers to a portion of DNA containing a repeating sequence of base pairs (micro- or minisatellite). The number of times a micro- or minisatellite repeats within specific loci can differ between individuals. These differences are used to distinguish donor DNA from recipient DNA. STR testing involves obtaining samples from the recipient and donor prior to transplant. Specific loci are compared to determine which loci contain STRs unique to the donor. After transplant, DNA is isolated from recipient samples. Donor-specific STRs are amplified by PCR techniques. The sample is then analyzed to determine the percent donor DNA present. |
| Amplified fragment length polymorphisms (AFLP) | A restriction fragment is a portion of DNA which has been cut out by an enzyme. AFLP testing begins by isolating DNA from the sample. Enzymes are used to cut the DNA at specific loci resulting in many unique restriction fragments. Many restrictions fragments are amplified using PCR techniques. The fragments are separated according to size by electrophoresis. The unique pattern of separation is used to identify the percent donor DNA present in the sample. Report AFLP testing using the VNTR/STR method option on the 2450 form. |

Chimerism Cell Types

<table>
<thead>
<tr>
<th>Cell Type</th>
<th>Description</th>
</tr>
</thead>
</table>
The peripheral blood or bone marrow sample has not been sorted or selected for a certain cell line.

- **Red blood cells**: Also known as RBCs or erythrocytes; carry the CD235a cell marker
- **Hematopoietic progenitor cells**: Includes CD34+ cells
- **Total mononuclear cells**: Total mononuclear cells would be a specimen containing only and both lymphocytes and monocytes
- **T cells**: Includes CD3+, CD4+, and / or CD8+ cells
- **B cells**: Includes CD19+ or CD20+ cells
- **Granulocytes**: Also known as polymorphonuclear leukocytes (PMNs, PMLs) and includes neutrophils, eosinophils, and basophils. Includes CD33+ cells
- **NK cells**: Includes CD56+ cells
- **Other**: Use this option to report cell types that do not fit in a category above.

### Section Updates:

<table>
<thead>
<tr>
<th>Question Number</th>
<th>Date of Change</th>
<th>Add/Remove/Modify</th>
<th>Description</th>
<th>Reasoning (If applicable)</th>
</tr>
</thead>
</table>

**Q85 – 107: Disease Assessment at the Time of Best Response to Infusion**

**Malignant Diseases Only**

Only complete Disease Assessment at the Time of Best Response to Infusion questions if the HCT being reported was given to treat a malignant disease. If the HCT being reported was given to treat a non-malignant disease, leave these questions blank. FormsNet3SM should enable / disable this section based on the primary disease reported on the Pre-TED Disease Classification Form (2402) Form. Contact the CIBMTR Center Support if you believe FormsNet3SM is incorrectly enabling / disabling these fields.

This section collects the data known as “best response to transplant.” The purpose of this section is to report the recipient’s best response to the planned course of the HCT. This includes response to any therapy given for post-HCT maintenance or consolidation and does not include response to treatment given for relapsed,
progressive or persistent disease. Best response is often achieved in the first 100 days. However, for some diseases such as multiple myeloma and CLL, the best response to HCT may take longer.

If the recipient relapses / progresses post-HCT and receives therapy for the disease relapse/progression, the response to that additional therapy should not be reported in this section. The best response prior to the relapse / progression should be reported. Reporting periods subsequent to that in which best response prior to the start of unplanned was reported will indicate that best response was previously reported.

**Reporting Complete Remission (CR) Post-HCT**

Complete remission (CR) criteria vary by disease and are outlined in the CIBMTR Forms Instructions Manual. Please refer to the appropriate disease response criteria section of the Forms Instructions Manual and review the criteria to report CR.

Tandem Transplants: For recipients receiving a tandem transplant, the best response to the prior transplant (i.e., HCT #1 of the tandem) depends on the pre-transplant disease status.

- If the recipient was in complete remission at the time of HCT #1 or achieved complete remission prior to HCT #2 of their tandem transplant, report the best response to transplant as “Continued complete remission (CCR)”.
- If the recipient was not in complete remission or did not achieve complete remission in response to HCT #1 prior to HCT #2 of their tandem transplant, either “Not in complete remission (NCR)” or “Not evaluated” would be appropriate options, however, ensure the best response to transplant and the current diseases status are answered consistently.

**Question 85:** Compared to the disease status prior to the preparative regimen, what was the best response to HCT since the date of the last report? (Include response to any therapy given for post-HCT maintenance or consolidation, but exclude any therapy given for relapsed, persistent, or progressive disease):

If the recipient was already in CR at the start of the preparative regimen, check **Continued complete remission (CCR)** and continue with question 108.

**Continued Complete Remission (CCR) should be reported for all patients who were already in CR at the start of the preparative regimen.**

If the recipient achieved CR post-HCT (excluding unplanned therapy), check **Complete remission (CR)** and continue with question 87.

If the recipient has not achieved a post-HCT CR to date, check **Not in complete remission** and continue with question 86.

If the recipient’s disease status was not evaluated post-HCT, check **Not evaluated** and continue with question 108. This option is **not** commonly used, as this would indicate...
that no tests (radiological, laboratory, or a clinical assessment) were performed to assess the CR status at any time during the reporting period.

If the recipient never achieved a post-transplant complete response and started unplanned therapy, given for relapsed, persistent, or progressive disease, in a previous reporting period, indicate Not evaluated.

Example 1: A recipient with neuroblastoma is not in complete remission prior to transplant, in the 100-day reporting period the recipient receives a tandem transplant. Between HCT 1 and HCT 2 the only disease assessment performed was a clinical evaluation. In this case either option would be appropriate to answer the best response to HCT as Not evaluated or Not in complete remission (NCR) and No disease detected but incomplete evaluation to establish CR.

Question 86: Specify disease status if not in complete remission
For recipients Not in complete remission, indicate whether clinical evidence of disease persisted on disease-specific assessments within the reporting period. If all assessments have shown resolution of disease, but not all assessments required to report complete remission have been completed, indicate No disease detected but incomplete evaluation to establish CR. This option is also appropriate for scenarios in which the recipient has not previously achieved a post-HCT CR but does not have any disease assessments performed within the reporting period. Indicate Disease detected if disease persists by any method of radiological or clinical assessment; persistence of abnormalities by molecular, cytogenetic, or flow cytometry assessments does not constitute “disease detected.”

Example 1: A recipient with multiple myeloma goes to transplant in VGPR, without a bone marrow showing < 5% plasma cells completed prior to transplant. Post-transplant serum and urine electrophoreses and immunofixations are negative. However, no bone marrow biopsy is performed within the 100-day reporting period. In this case, Not in complete remission should be selected for best response to HCT, and No disease detected by incomplete evaluation to establish CR for specifying the disease status if not in complete remission data field.

Example 2: A recipient with AML goes to transplant in primary induction failure. Post-transplant, they recover their counts, but had circulating blasts noted on differential. They expire due to persistent disease with their last CBC performed on their date of death showing circulating blasts. In this case, Not in complete remission should be selected for the best response to HCT, and Disease detected for specifying the disease status if not in complete remission data field.

Example 3: Similar to example 2, a recipient with AML goes to transplant in primary induction failure. They expire on D+11 due to infection and had not
engrafted as of that date. Their last CBC showed a WBC of $0.5 \times 10^9/L$ with no blasts detected on their differential. A bone marrow biopsy was not performed between transplant and the date of death. In this case, **Not in complete remission** should be selected for the best response to HCT, and **No disease detected by incomplete evaluation to establish CR** for specifying the disease status if not in complete remission data field.

**Question 87: Was the date of best response previously reported?**
Indicate whether complete remission was reported in a previously reporting period; if Yes, continue with question 108. This question does not apply if the best response is **Not in complete remission**.

**Question 88: Date assessed**
Report the date complete remission was achieved. This date should fall after transplant but before or on the date of contact for the current reporting period. This should reflect the date of specimen collection or imaging for the latest assessment required to fulfill complete remission criteria for the recipient’s transplant disease.

**Disease Assessment at Time of Best Response**

Questions 89 – 107 refer to disease assessments performed at the time of best response (question 85). The following guidelines should be used to determine whether testing was performed at the time of best response:

If the recipient’s best response is **Not in Complete Remission**, report the latest assessment performed during the reporting period. If the recipient has started treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease), report the latest assessment prior to the initiation of therapy.

If the recipient’s best response is **Complete Remission**, report testing performed closest to the date of best response (questions 88) and within the time windows in the Disease Assessment Time Windows table.

**Disease Assessment Time Windows**

<table>
<thead>
<tr>
<th>Follow-Up Form</th>
<th>Approximate Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>100 Day</td>
<td>+/- 15 days of date of best response (question 78)</td>
</tr>
<tr>
<td>6 Month</td>
<td>+/- 15 days of date of best response (question 78)</td>
</tr>
<tr>
<td>Annual</td>
<td>+/- 30 days of date of best response (question 78)</td>
</tr>
</tbody>
</table>

**Disease Assessment Reporting Scenarios:**

**A.** A recipient receives a transplant on 1/1/2015 for multiple myeloma in partial remission. Prior to HCT, FISH testing detects an IGH rearrangement associated with the recipient’s primary disease. During the 100 day reporting
period, the recipient achieves a very good partial remission. FISH testing is only performed on 2/1/2015 is positive for the previously detected IGH rearrangement. The 100 day date of contact is 4/15/2015. In this case, the center would report the recipient was “Not in Complete Remission” on the 100 Day Post-TED Form. The center would report FISH testing was performed on 2/1/2015. When the best response is “Not in Complete Remission” report the most recent testing performed during the reporting period (assuming treatment was not started for relapsed, progressive, or persistent disease during the reporting period – see Scenario B).

B. A recipient receives a transplant on 1/1/2015 for multiple myeloma in partial remission. Prior to HCT, FISH testing detects an IGH rearrangement associated with the recipient’s primary disease. During the 100 day reporting period, the recipient has disease progression and starts treatment on 3/1/2015. FISH testing is performed on 2/1/2015 and 3/15/2015. Both tests are positive for the previously detected IGH rearrangement. The 100 day date of contact is 4/15/2015. In this case, the center would report the recipient was “Not in Complete Remission” on the 100 Day Post-TED Form. The center would report FISH testing was performed on 2/1/2015. When the best response is “Not in Complete Remission” report the most recent testing performed during the reporting period and prior to treatment for relapsed, progressive or persistent disease.

Note: For all subsequent reporting periods, the center would report “Not Evaluated” for the best response to HCT and skip to the Post-HCT Therapy section of the form. If treatment was started in a prior reporting period, the center is not able to report and assessments performed during the reporting period and prior to treatment

C. A recipient receives a transplant on 1/1/2015 for AML in primary induction failure. Prior to HCT, molecular testing confirms the recipient’s disease is FLT3 positive. On 2/1/2015, the recipient achieves a hematologic remission, but FLT3 is not tested at that time. Later, on 2/10/2015, molecular testing is performed and confirms the recipient is FLT3 negative. In this case, the center would report the recipient achieved a CR on 2/1/2015 on the 100 Day Post-TED Form. The center would report molecular testing was performed at the time of best response as testing was done within 15 days of 2/1/2015.

D. A recipient receives a transplant on 1/1/2015 for AML in primary induction failure. Prior to HCT, molecular testing confirms the recipient’s disease is FLT3 positive. On 2/1/2015, the recipient achieves a hematologic remission, but FLT3 is not tested at that time. Later, on 3/1/2015, molecular testing is performed and confirms the recipient is FLT3 negative. In this case, the center would report the recipient achieved a CR on 2/1/2015 on the 100 Day Post-TED Form. The center would report no molecular testing was performed.
at the time of best response as testing was not done within 15 days of 2/1/2015.

**E.** A recipient receives a transplant on 1/1/2015 for NHL in stable disease. During the 100 Day reporting period, a PET / CT was performed on Day 60, confirming stable disease but then on Day 95, another PET / CT was performed and showed progression. As a result, therapy for progression began on Day 100. The best response to HCT for the Day 100 reporting period would be reported as “Not in complete remission – disease detected” and report “Yes,” radiologic assessments were performed with the Day 60 PET / CT as this is the most recent scan prior to progression.

**F.** A recipient receives a transplant on 1/1/2020 for IgA Kappa Multiple Myeloma in stable disease. During the 100 Day reporting period, the first set of myeloma labs on Day 29, 1/30/2020, show progressive disease. Myeloma labs repeated on Day 60 and Day 100 also showed disease progression. As a result, therapy is planned to be given, starting in the 6-month reporting period, on Day 110. The best response to HCT for the Day 100 reporting period would be reported as “Not in complete remission – disease detected” and report “Yes,” clinical / hematologic assessments were performed with the Day 100 myeloma labs, as this is the most recent testing in the reporting period. In cases where the first assessment post-HCT shows progression, report the last assessment prior to the start of treatment. If treatment doesn’t start until the next reporting period, report the last assessment in the current reporting period.

**Molecular**

Questions 89 – 91 are intended to capture molecular abnormalities identified by molecular methods. Additional testing methods, such as FISH, may identify molecular marker results but should not be reported in the molecular section of the Post-TED (2450) Form. Abnormalities identified by karyotyping, FISH, or microarray should only be reported in the cytogenetic section of the Post-TED (2450) Form.

**Question 89: Was the disease status assessed by molecular testing (e.g. PCR)?**
Molecular assessment involves determining whether a molecular marker for the disease exists in the blood or bone marrow. Molecular assessment is the most sensitive method of detection and can indicate known genetic abnormalities associated with the disease for which the HCT was performed. Molecular assessments include polymerase chain reaction (PCR) amplification to detect single specific disease markers; however, molecular methods are evolving and now include chromosomal microarray / chromosomal genomic array, Sanger sequencing, and next generation sequencing (e.g., Illumina, Roche 454, Proton / PGM, SOLiD).
Report **Not applicable** if molecular studies were never performed or have never shown abnormalities associated with the recipient’s primary transplant disease.

Once an assessment is positive for disease, **Not applicable** will never be an appropriate response.

Report **No** if molecular studies were not performed during the reporting period.

If the recipient’s best response is **Not in Complete Remission**, report the latest assessment performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease). If testing was not performed prior to the initiation of treatment, report **No** and go to question 92.

If the recipient’s best response is **Complete Remission**, report testing performed closest to the date of best response (question 85) and within the time windows in the Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report **No** and go to question 92.

**Question 90: Date assessed**

If the best response is **Complete remission**, report the date of testing performed nearest the date of best response and prior to relapse or progression, if applicable.

If the best response is **Not in complete remission**, report the date of the most recent molecular testing performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease, if applicable. If no treatment for relapsed, progressive, or persistent disease was given, report the date of the most recent molecular assessment performed within approximately 30 days of the follow-up date.

Report the date of specimen collection for molecular disease assessment. If exact date is not known, refer to General Instructions, General Guidelines for Completing Forms for information about reporting partial or unknown dates.

**Question 91: Was disease detected?**

Report whether the recipient’s primary disease was detected by molecular testing on the date reported in question 89. In order to be considered positive for disease, the assay must detect a number of copies of the molecular marker exceeding the threshold for sensitivity of the assay, for a quantitative study. However, do note that presence of only a single marker among numerous tested is sufficient to indicate disease detected.

**Flow Cytometry**

**Question 92: Was the disease status assessed via flow cytometry?**
Flow cytometry is a technique that can be performed on blood, bone marrow, or tissue preparations where cell surface markers can be quantified on cellular material. This allows for the detection of abnormal cell populations for some diseases.

Report Not applicable if flow cytometry was never performed or have never shown abnormalities associated with the recipient’s primary transplant disease.

Once an assessment is positive for disease, Not applicable will never be an appropriate response.

Report No if flow cytometry was not performed during the reporting period.

If the recipient’s best response is Not in Complete Remission, report the latest assessment performed during the reporting period and prior to any treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease). If testing was not performed prior to the initiation of treatment, report No and go to question 95.

If the recipient’s best response is Complete Remission, report testing performed closest to the date of best response (question 85) and within the time windows in the Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report No and go to question 95.

**Question 93: Date assessed**

If the best response is Complete remission, report the date of testing performed nearest the date of best response and prior to relapse or progression, if applicable.

If the best response is Not in complete remission, report the date of the most recent flow cytometry testing performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease, if applicable. If no treatment for relapsed, progressive, or persistent disease was given, report the date of the most recent flow cytometry performed within approximately 30 days of the follow-up date.

Report the date of specimen collection for flow cytometry assessment. If exact date is not known, refer to General Instructions, General Guidelines for Completing Forms, for information about reporting partial or unknown dates.

**Question 94: Was disease detected?**

Report whether the recipient’s primary disease was detected by flow cytometry on the date reported in question 93. Report Yes if an abnormal cell population associated with the recipient’s primary transplant disease was detected regardless of the sensitivity of the flow cytometry panel performed; this means an abnormal cell population detected by MRD flow cytometry would be reported in the same way as an abnormal cell population detected by a standard flow cytometry assay.
Cytogenetic Testing (Karyotyping or FISH)

Question 95: Was the disease status assessed by cytogenetic testing (karyotyping or FISH)?

Cytogenetic studies involve the study of chromosomes, typically through one of two methods: karyotyping or fluorescence in situ hybridization (FISH). Blood, bone marrow, or tissue preparations may be tested by either of these two methods. Karyotyping is both less sensitive and less specific than FISH testing; FISH studies identify only abnormalities detectable by the employed probe set and cannot provide information about the presence or absence of chromosomal abnormalities or markers outside the specific probe set utilized.

Report Not applicable if cytogenetic studies were never performed or have never shown abnormalities associated with the recipient’s primary transplant disease.

Once an assessment is positive for disease, Not applicable will never be an appropriate response.

Report No if cytogenetic studies were not performed during the reporting period.

If the recipient's best response is Not in Complete Remission, report the latest assessment performed during the reporting period and prior to any treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease). If testing was not performed prior to the initiation of treatment, report No and go to question 102.

If the recipient's best response is Complete Remission, report testing performed closest to the date of best response (question 81) and within the time windows in the Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report No and go to question 102.

Question 96: Was the disease status assessed via FISH?

FISH XX/XY probe sets are not considered relevant to disease assessment and should not be reported in the disease assessment section.

Report Not applicable if FISH studies were never performed or have never shown abnormalities associated with the recipient’s primary transplant disease.

Once an assessment is positive for disease, Not applicable will never be an appropriate response.

Report No if FISH studies were not performed during the reporting period.

If the recipient’s best response is Not in Complete Remission, report the latest assessment performed during the reporting period and prior to any treatment for
relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease). If testing was not performed prior to the initiation of treatment, report No and go to question 99.

If the recipient’s best response is Complete Remission, report testing performed closest to the date of best response (question 85) and within the time windows in the Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report No and go to question 99.

**Question 97: Date assessed**
If the best response is Complete remission, report the date of testing performed nearest the date of best response and prior to relapse or progression, if applicable.

If the best response is Not in complete remission, report the date of the most recent FISH testing performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease, if applicable. If no treatment for relapsed, progressive, or persistent disease was given, report the date of the most recent FISH assessment performed within approximately 30 days of the follow-up date.

Report the date of specimen collection for FISH assessment. If exact date is not known, refer to General Instructions, General Guidelines for Completing Forms, for information about reporting partial or unknown dates.

**Question 98: Was disease detected?**
Report whether the recipient’s primary disease was detected by FISH testing on the date reported in question 97.

**Question 99: Was the disease status assessed via karyotyping?**
Report Not applicable if karyotyping was never performed or have never shown abnormalities associated with the recipient’s primary transplant disease.

Once an assessment is positive for disease, Not applicable will never be an appropriate response.

Report Not if karyotyping was not performed during the reporting period.

If the recipient’s best response is Not in Complete Remission, report the latest assessment performed during the reporting period and prior to any treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease). If testing was not performed prior to the initiation of treatment, report No and go to question 102.

If the recipient’s best response is Complete Remission, report testing performed closest to the date of best response (question 85) and within the time windows in the
Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report No and go to question 102.

**Question 100: Date assessed**
If the best response is **Complete remission**, report the date of testing performed nearest the date of best response and prior to relapse or progression, if applicable.

If the best response is **Not in complete remission**, report the date of the most recent karyotype testing performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease, if applicable. If no treatment for relapsed, progressive, or persistent disease was given, report the date of the most recent karyotype performed within approximately 30 days of the follow-up date.

Report the date of specimen collection for karyotyping. If exact date is not known, refer to General Instructions, General Guidelines for Completing Forms, for information about reporting partial or unknown dates.

**Question 101: Was disease detected?**
Report whether the recipient’s primary disease was detected by karyotyping on the date reported in question 97. Do not include clinically insignificant polymorphism, or chromosomal abnormalities of no known significance, as disease detected; this includes anomalies such as age-dependent loss of the chromosome Y.

**Radiologic**

**Question 102: Was the disease status assessed by radiological assessment (e.g. PET, MRI, CT)**
Radiologic assessments are imaging techniques used to assess disease response to transplant, typically for lymphomas or solid tumors, though valuable in some less common presentations of disease, such as leukemia cutis. Imaging techniques used to evaluate disease response typically include PET, CT, or MIBG, but may include x-ray, skeletal survey, or ultrasound in some cases.

Report **Not applicable** if radiological assessments were never performed or have never shown abnormalities associated with the recipient’s primary transplant disease.

Once an assessment is positive for disease, **Not applicable** will never be an appropriate response.

Report **No** if radiological assessments were not performed during the reporting period.

If the recipient’s best response is **Not in Complete Remission**, report the latest assessment performed during the reporting period and prior to any treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual
If testing was not performed prior to the initiation of treatment, report **No** and go to question 105.

**If the recipient’s best response is Complete Remission**, report testing performed closest to the date of best response (questions 85) and within the time windows in the Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report **No** and go to question 105.

**Question 103: Date assessed**
If the best response is **Complete remission**, report the date of the assessment performed nearest the date of best response and prior to relapse or progression, if applicable.

If the best response is **Not in complete remission**, report the date of the most recent radiologic testing performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease, if applicable. If no treatment for relapsed, progressive, or persistent disease was given, report the date of the most recent radiological assessment performed within approximately 30 days of the follow-up date.

Report the date of radiological assessment. For recipients with **Complete remission** reported in question 85, this may match the date CR was achieved reported in question 88 for recipients with lymphomas, solid tumors, or other diseases with imaging criteria for reporting CR. If exact date is not known, refer to General Instructions, **General Guidelines for Completing Forms**, for information about reporting partial or unknown dates.

**Question 104: Was disease detected?**
Report whether the recipient’s primary disease was detected by radiologic assessment on the date reported in question 103.

**Clinical / Hematologic**

**Question 105: Was the disease status assessed by clinical / hematologic assessment?**
Clinical / hematologic disease assessments are the least sensitive method of disease detection. Examples include circulating blasts in the bloodstream for AML, and enlargement of a malignant mass for lymphoma or a solid tumor on physical examination. Every recipient who has an evaluation by a physician has a “clinical” assessment. Do not include radiologic or imaging assessments when answering this question.

**If the recipient’s best response is Not in Complete Remission**, report the latest assessment performed during the reporting period and prior to any treatment for relapsed, progressive, or persistent disease (excluding treatment for minimal residual disease).
disease). If testing was not performed prior to the initiation of treatment, report No and go to question 108.

If the recipient’s best response is Complete Remission, report testing performed closest to the date of best response (question 85) and within the time windows in the Disease Assessment Time Windows table. If testing was not performed within the applicable time window, report No and go to question 108.

**Question 106: Date assessed**
If the best response is Complete remission, report the date of the assessment performed nearest the date of best response and prior to relapse or progression, if applicable. This will likely match the date CR was achieved reported in question 85, since complete remission criteria generally require clinical or hematologic assessment to confirm.

If the best response is Not in complete remission, report the date of the most recent clinical / hematologic testing performed during the reporting period and prior to treatment for relapsed, progressive, or persistent disease, if applicable. If no treatment for relapsed, progressive, or persistent disease was given, report the date of the most disease-specific testing performed within approximately 30 days of the follow-up date.

If exact date is not known, refer to General Instructions, *General Guidelines for Completing Forms*, for information about reporting partial or unknown dates.

**Question 107: Was disease detected?**
Report whether clinical / hematologic abnormalities associated with the primary disease were detected. In general, if the clinical/hematologic assessment date is that same as that reported in question 85, for recipients achieving complete remission in the reporting period, the answer to this question should be No.

**Section Updates:**

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**Q108 – 112: Post-Infusion Therapy**

**Malignant Diseases Only**
Only complete questions 108 – 112 if the HCT being reported was given to treat a malignant disease. If the HCT being reported was given to treat a non-malignant disease, leave questions 108-112 blank. FormsNet3SM should enable / disable this
Report therapy given since the date of last report for reasons other than relapse, persistent, or progressive disease. This may include maintenance and consolidation therapy as well as treatment for minimal residual disease. Do not report any therapy given for relapse, persistent, or progressive disease.

**Question 108: Was therapy given since the date of the last report for reasons other than relapse, persistent, or progressive disease? (Include maintenance and consolidation therapy)**

Indicate whether therapy was given during the reporting period for maintenance or consolidation; this therapy may have been specifically planned as part of the original transplant protocol or determined after transplant. Do not include therapy given for relapse, persistent, or progressive disease. Any post-transplant therapy included as part of the initial transplant protocol should be reported in this area of the form.

**Question 109: Specify therapy (check all that apply)**

Indicate which therapies were given since the date of the last report for reasons other than relapse, persistent, or progressive disease.

**Systemic therapy**: Refers to a delivery mechanism where a therapeutic agent is delivered orally or intravenously, enters the bloodstream, and is distributed throughout the body. Indicate whether systemic therapy was given during the reporting period for reasons other than relapse, persistent, or progressive disease and report the systemic therapy given in question 110.

**Radiation**: Radiation therapy uses high-energy radiation to kill cancer cells. External beam radiation is one of the more frequently used types of radiation. In this method, a beam of radiation is delivered to a specific part of the body, such as the mediastinum. Radiation may be planned if bulky disease was present just prior to transplant for a recipient with lymphoma or a solid tumor. Indicate whether radiation therapy was given during the reporting period for reasons other than relapse, persistent, or progressive disease and go to question 113.

**Cellular therapy**: Cellular therapy refers to the infusion of human or animal derived cells, which may or may not be modified or processed to achieve a specific composition. Examples include CAR T-cell, NK cell, and mesenchymal cell infusions as well as donor cellular infusions. Indicate **Yes** if the recipient received any form of cellular therapy for reasons other than relapse, persistent, or progressive disease; hematopoietic cell transplantation should not be reported as cellular therapy. Indicate whether a cellular therapy was infused during the reporting period for reasons other than relapse, persistent, or progressive disease and go to question 113.
Blinded randomized trial: A blinded, randomized trial refers to a research treatment protocol in which the participant is assigned to the control arm or investigational group, and the researcher or clinician is not informed whether the subject is receiving the placebo or standard of care versus the investigational therapy. This makes it impossible to report agents or therapies the recipient is receiving. Indicate whether the recipient is receiving therapy on a randomized, blinded clinical trial during the reporting period for reasons other than relapse, persistent, or progressive disease and go to question 113.

Other therapy: Indicate whether the recipient received additional therapy for reasons other than relapsed, persistent, or progressive disease which does not fit into the previous categories. Examples may include intrathecal therapy or surgery. Specify the other therapy given in question 112.

Questions 110 – 111: Specify systemic therapy (check all that apply)
Systemic therapy agents and treatment regimens vary based on disease, prognosis, and protocol. Treatment may consistent of one or multiple drugs, and may be given in an inpatient or outpatient setting; additionally, drugs may be administered on a single day, over consecutive days, or continuously.

Form options are arranged alphabetically. Indicate which systemic therapy agents were administered during the current reporting period for reasons other than relapse, persistent, or progressive disease. If the recipient received a therapeutic agent that is not listed, select Other systemic therapy and specify the systemic. Only report maintenance or consolidation therapy for the primary disease for infusion and do not include drugs like bisphosphonates, such as Zometa, given to recipients with multiple myeloma to improve bone strength.

Steroids Administered Post-HCT
Previously, steroids given for reasons other than relapsed, persistent, or progressive disease were reported in the Other systemic therapy section of the Post-TED (2450) Form. We no longer capture steroids (e.g. dexamethasone) on our Post-TED (2450) Form and they should not be reported here.

Question 112: Specify other therapy
Specify other therapy the recipient received additional therapy for reasons other than relapsed, persistent, or progressive disease which does not fit into the previous form categories. Examples may include intrathecal therapy or surgery.

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DRAFT Instructions for Post-TED
CIBMTR Form 2450

Q113 – 123: Relapse or Progression Post-Infusion

Malignant Diseases Only

Only complete questions 113 – 115 if the HCT being reported was given to treat a malignant disease. If the HCT being reported was given to treat a non-malignant disease, leave questions 113 – 115 blank. Questions 116 – 123 (intervention for relapsed, persistent, or progressive disease) must be completed regardless of disease type. FormsNet3SM should enable / disable this section based on the primary disease reported on the Pre-TED Disease Classification (2402) Form. Contact the CIBMTR Center Support if you believe FormsNet3SM is incorrectly enabling / disabling these fields.

Report if the recipient has experienced a clinical/hematologic relapse or progression post-HCT. If the relapse or progression was detected in a previous reporting period indicate that and continue on. If the first clinical/hematologic relapse occurred since the date of the last report, indicate the date it was first detected in this reporting period.

Question 113: Did the recipient experience a clinical / hematologic relapse or progression post-HCT?
Clinical / hematologic assessment is the least sensitive method of disease detection. Examples include circulating blasts in the bloodstream for AML, or enlargement of a malignant mass for lymphoma or a solid tumor. Every recipient who has an evaluation by a physician has a “clinical” assessment. Include radiographic evidence of relapse or progression as clinical/hematologic relapse or progression. Disease specific criteria for establishing relapse or progression are published as part of the CIBMTR Forms Instructions Manual. If the recipient dies, and the relapse or progression of disease is discovered by autopsy, the date of assessment should be reported as the date of death, not the autopsy date.

If clinical / hematologic evidence of relapse / progressive disease was found at any time post-transplant, check Yes.

If clinical / hematologic evidence of relapse / progressive disease was not found at any time post-transplant, check No and continue with question 116.

Question 114: Was the date of the first clinical / hematologic relapse or progression previously reported?

Only the date of first clinical / hematologic relapse or progression post-transplant needs to be reported. Therefore, if the recipient experienced clinical/hematologic relapse or progression in a prior reporting period and it was reported on the prior form, report Yes.
and continue with question 124. If this is the report of first instance of clinical / hematologic relapse or progression, indicate No.

**Question 115: Date first seen**
Indicate the date relapse / progressive disease was determined by clinical / hematological evaluation. If exact date is not known, refer to General Instructions, General Guidelines for Completing Forms, for information about reporting partial or unknown dates.

**Question 116: Was intervention given for relapsed, persistent or progressive disease since the date of last report?**
Indicate whether therapy was given during the reporting period for persistent or relapsed / progressive disease. Do not include therapy given for maintenance or planned post-transplant consolidation. Any post-transplant therapy included as part of the initial transplant protocol should not be reported in this area of the form. See the intervention reporting scenarios provided below for further clarification.

**Question 117: Specify reason for which intervention was given**
Indicate whether therapy was given for persistent or relapsed / progressive disease. If therapy continued from a prior reporting period and a new therapy was started for a different reason during the current reporting period, report the reason the new therapy was started. See the intervention reporting scenarios provided below for further clarification.

**Question 118: Specify the method(s) of detection for which intervention was given (check all that apply)**
Indicate the methods detecting the reason for which therapy for persistent disease or relapsed / progressive disease was given (as reported in question 117). Indicate all methods of disease assessment in which disease was detected; given that the assessment was performed prior to the start of the intervention(s) and was consistent with the rationale reported in question 117. There may be some cases for which an assessment by a particular method was last performed in the prior reporting period but was still consistent with the justification reported in question 117; in this case, the method of disease assessment should be indicated.

For example, in the 100-day reporting period, the last cytogenetic assessment detected a new abnormality associated with the recipient’s primary transplant disease. In this case, monosomy 7 was identified on a peripheral blood sample for a recipient transplanted for AML in CR1 with normal cytogenetics prior to transplant. In the 6-month reporting period, relapse was detected in the bone marrow morphology (clinical assessment) and concurrent flow cytometry (flow cytometry) and therapy was initiated for relapsed / progressive disease. In this case, each of these methods should be indicated on the Post-TED (2450) Form in the 6-month reporting period.
If multiple therapies were given during the reporting period for different reasons (e.g., the recipient initially receives treatment for persistent disease and subsequently receives different treatment for progressive disease during the same reporting period), indicate any methods of detection confirming the reason in question 117. See the intervention reporting scenarios provided below for further clarification.

If assessment by that method was not performed or was performed and not consistent with the reason for which intervention was given reported in question 117, do not indicate it in this question.

See below for definitions and examples of each method of detection:

- **Clinical/hematologic**: Clinical / hematologic assessment is the least sensitive method of disease detection. Examples include circulating blasts in the bloodstream for AML, or enlargement of a malignant mass for lymphoma or a solid tumor. Every recipient who has an evaluation by a physician has a “clinical” assessment. Examples of clinical/hematologic assessments include: bone marrow biopsy / morphologic evaluation, complete blood count, serum protein electrophoresis, etc.

- **Radiologic (e.g., PET, MRI, CT)**: Radiologic assessments are imaging techniques used to assess disease response. Imaging techniques used to evaluate disease response typically include PET, CT, or MIBG, but may include x-ray, skeletal survey, or ultrasound in some cases.

- **Cytogenetic**: Cytogenetic studies involve the study of chromosomes, typically through one of two methods: karyotyping or fluorescence in situ hybridization (FISH). Blood, bone marrow, or tissue preparations may be tested by either of these two methods. Karyotyping is both less sensitive and less specific than FISH testing; FISH studies identify only abnormalities detectable by the employed probe set and cannot provide information about the presence or absence of chromosomal abnormalities or markers outside the specific probe set utilized.

- **Flow cytometry**: Flow cytometry is a technique that can be performed on blood, marrow, or tissue preparations where the cell surface markers can be quantified on cellular material. This allows for the detection of abnormal cell populations for some diseases. Flow cytometry may also be referred to as immunophenotyping.

- **Disease specific molecular marker**: Molecular assessment involves determining whether a molecular marker for the disease exists in the blood or bone marrow. Molecular assessment is the most sensitive method of detection and can indicate known genetic abnormalities associated with the disease for which the HCT was performed.

**Question 119: Date intervention started**
Report the date therapy was started for the reason specified in question 117; if multiple instances, cycles, or lines of therapy are administered, report the date of the first treatment. If treatment was started in a prior reporting period and continues into the
current reporting period, report the original therapy start date (prior to the start of the current reporting period) and override the validation error in FormsNet3SM using the code “verified correct.” If therapy was stopped in a prior reporting period and restarted (or a new therapy was started) during the current reporting period, report the earliest date treatment was administered during the current reporting period. See the intervention reporting scenarios provided below for further clarification.

**Intervention Reporting Scenarios**

A. A recipient with NHL in complete remission at the time of HCT has a relapse during the 100 day reporting period. Relapse was detected by a PET scan and a lymph node biopsy. Following these assessments, rituximab was started on 5/1/2016. The disease did not respond to this therapy prompting a switch to brentuximab on 6/1/2016. The 100 Day date of contact is 6/15/2016.

*100 Day Post-TED Form:*

Q116: Report “Yes” to indicate therapy was given for relapsed disease during this reporting period.
Q117: Report “Relapsed / progressive disease.”
Q118: Check the boxes to indicate that disease was detected by both clinical/hematologic (lymph node biopsy) and radiological (PET Scan) assessments. All other methods of detection must be left blank.
Q119: Report “5/1/2016” to reflect the date of the first treatment given for relapsed disease.
Q120 – 123: Report both rituximab and brentuximab as treatments for relapsed disease given during the reporting period.

B. A recipient with multiple myeloma in VGPR at the time of HCT was started on maintenance lenalidomide during the six-month reporting period. Later in the reporting period, progression was detected by serum protein electrophoresis on 9/15/2014 and so the recipient stopped lenalidomide and started bortezomib as well as dexamethasone on 9/20/2014. The recipient continued bortezomib and dexamethasone treatment into the one-year reporting period.

*Six Month Post-TED Form:*

Q116: Report “Yes” to indicate therapy was given for progressive disease during this reporting period.
Q117: Report “Relapsed / progressive disease.”
Q118: Check the box to indicate that disease was detected by clinical/hematologic (serum protein electrophoresis) assessment. All other methods of detection must be left blank.
Q119: Report “9/20/2014” to reflect the date of the first treatment given for progressive disease.
Q120 – 123: Report bortezomib as treatment for progressive disease given during the reporting period. Dexamethasone is no longer captured on the Post-TED (2450) Form. The lenalidomide therapy should not be reported in this section of the form. This medication was given as maintenance therapy and will therefore be reported under Post-HCT Therapy.

One Year Post-TED Form:

Q115 – 123: These questions will be disabled in FormsNet3. Starting with Revision 5 of the Post-TED (2450), therapy given for relapsed or progressive disease will only be captured in the reporting period in which treatment first started.

C. A recipient with multiple myeloma in PR at the time of HCT was started on lenalidomide during 100-day reporting period (started 3/15/2012) due to persistent disease (detected by serum electrophoresis testing). This treatment was not planned and was given due to an unsatisfactory disease response to HCT. Thirty days after lenalidomide was started, a karyotype assessment confirmed persistent cytogenetic abnormalities present in a bone marrow sample. Lenalidomide was continued into the six-month reporting period, during which, there was disease progression (detected by serum electrophoresis). Lenalidomide was stopped and carfilzomib was started on 5/30/2012. By the end of the six-month reporting period, the recipient achieved a complete remission in response to carfilzomib and was switched to a lower maintenance dose of carfilzomib which was continued into the one-year reporting period.

100 Day Post-TED Form:

Q116: Report “Yes” to indicate therapy was given for persistent disease during this reporting period.
Q117: Report “Persistent disease.”
Q118: Check the box to indicate that disease was detected by clinical/hematologic (serum protein electrophoresis) assessment. All other methods of detection must be left blank. The karyotype test would not be reported as a method of detection since it was performed after treatment was started and, therefore, did not inform the decision to start lenalidomide.
Q119: Report “3/15/2012” to reflect the date of the first treatment for persistent disease.
Q120 – 123: Report lenalidomide as the only treatment given during the reporting period.

Six Month Post-TED Form:

Q116: Report “Yes” to indicate therapy was given for persistent and progressive disease during this reporting period.
Q117: Report “relapsed / progressive disease.” If therapy continued from a prior reporting period and a new therapy was started for a different reason during the current reporting period, report the reason the new therapy was started.

Q118: Check the box to indicate that disease was detected by clinical/hematologic (serum protein electrophoresis) assessment. All other methods of detection must be left blank.

Q119: Report “5/30/2012” to reflect the date of the first treatment for progressive disease.

Q120 – 123: Report the lenalidomide and carfilzomib as treatments received during the reporting period.

One Year Post-TED Form:

Q116: Report “No” to indicate therapy was not given for persistent or relapsed / progressive disease during this reporting period. The lower dose carfilzomib given as maintenance (to keep the recipient in CR) must be reported in the Post-HCT Therapy Section of the Post-TED Form. Reporting “No” for question 116 will disable questions 117 – 123.

Question 120: Specify therapy (check all that apply)

Indicate which therapies were given since the date of the last report for relapsed, persistent, or progressive disease.

**Systemic therapy:** refers to a delivery mechanism where a therapeutic agent is delivered orally or intravenously, enters the bloodstream, and is distributed throughout the body. Indicate whether systemic therapy was given during the reporting period for relapsed, persistent, or progressive disease and report the systemic therapy given.

**Radiation:** Radiation therapy uses high-energy radiation to kill cancer cells. External beam radiation is one of the more frequently used types of radiation. In this method, a beam of radiation is delivered to a specific part of the body, such as the mediastinum. Radiation may be planned if bulky disease was present just prior to transplant for a recipient with lymphoma or a solid tumor. Indicate whether radiation therapy was given during the reporting period for relapsed, persistent, or progressive disease.

**Cellular therapy:** Cellular therapy refers to the infusion of human or animal derived cells, which may or may not be modified or processed to achieve a specific composition. Examples include CAR T-cell, NK cell, and mesenchymal cell infusions as well as donor cellular infusions. Indicate **Yes** if the recipient received any form of cellular therapy for relapse, persistent, or progressive disease; hematopoietic cell transplantation should not be reported as cellular therapy. Indicate whether a cellular therapy was infused during the reporting period for relapsed, persistent, or progressive disease.
**Blinded randomized trial:** A blinded, randomized trial refers to a research treatment protocol in which the participant is assigned to the control arm or investigational group, and the researcher or clinician is not informed whether the subject is receiving the placebo or standard of care versus the investigational therapy. This makes it impossible to report agents or therapies the recipient is receiving. Indicate whether the recipient is receiving therapy on a randomized, blinded clinical trial during the reporting period for relapsed, persistent, or progressive disease.

**Other therapy:** Indicate whether the recipient received additional therapy for relapsed, persistent, or progressive disease which does not fit into the previous categories. Examples may include intrathecal therapy or surgery. Specify the other therapy.

**Questions 121 – 122: Specify systemic therapy (check all that apply)**
Systemic therapy agents and treatment regimens vary based on disease, prognosis, and protocol. Treatment may consist of one or multiple drugs, and may be given in an inpatient or outpatient setting; additionally, drugs may be administered on a single day, over consecutive days, or continuously.

Form options are arranged alphabetically. Indicate which systemic therapy agents were administered during the current reporting period for reasons other than relapse, persistent, or progressive disease. If the recipient received a therapeutic agent that is not listed (e.g. cyclophosphamide), **Chemotherapy** should be selected. If the recipient received a therapeutic agent, other than chemotherapy, that is not listed, select **Other systemic therapy**, and specify the systemic therapy.

**NOTE: Steroids Administered Post-HCT**
Previously, steroids given for relapsed, persistent, or progressive disease were reported in the “other systemic therapy” section of the Post-TED (2450) Form. We no longer capture steroids (e.g. dexamethasone) on our Post-TED (2450) Form and they should not be reported here.

**Question 123: Specify other therapy**
Specify other therapy the recipient received additional therapy for reasons other than relapsed, persistent, or progressive disease which does not fit into the previous form categories. Examples may include intrathecal therapy or surgery.

**Section Updates:**
**Q124 – 127: Current Disease Status**

**Malignant Diseases Only**

Only complete the Current Disease Status section if the HCT being reported was given to treat a malignant disease. If the HCT being reported was given to treat a non-malignant disease, leave these questions blank. FormsNet3SM should enable / disable this section based on the primary disease reported on the Pre-TED Disease Classification (2402) Form. Contact the CIBMTR Center Support if you believe FormsNet3SM is incorrectly enabling / disabling these fields.

Tandem Transplants: For recipients receiving a tandem transplant, the current disease status prior to HCT #2 of the tandem depends on the pre-transplant disease status and the best response to the prior transplant (i.e., HCT #1 of the tandem).

- If the recipient was in complete remission at the time of HCT #1 or achieved complete remission prior to HCT #2 of their tandem transplant, the current disease status should be reported as “Complete remission (CR)” (given there is no evidence of relapse / progression disease based on labs / clinical assessments between the tandem HCTs).
- If the recipient was not in complete remission or did not achieve complete remission in response to HCT #1 prior to HCT #2 of their tandem transplant, either “Not in complete remission (NCR)” or “Not evaluated” would be appropriate options; however, ensure the best response to transplant and the current diseases status are answered consistently.

**Question 124: What is the current disease status?**

Indicate the disease status of the primary transplant disease as of the last evaluation in the reporting period. Complete remission (CR) criteria vary by disease and are outlined in the CIBMTR Forms Instructions Manual. If the recipient achieves CR or continues in CR at the time of last evaluation in the reporting period, indicate **Complete remission (CR)**. If the recipient is not in CR due to presence of disease on last evaluation in the reporting period or an incomplete evaluation that does not allow for reporting CR, indicate **Not in complete remission**. If the recipient’s disease status was not evaluated post-HCT, check **Not evaluated** and submit the form. This option is **not** commonly used, as this would indicate that **no tests** (radiological, laboratory, or a clinical assessment) were performed to assess the CR status at **any time** during the reporting period.

The center does not need to repeat all disease-specific assessments (biopsies, scans, labs) each reporting period in order to complete current disease status data fields. Once a particular disease status is achieved, the center can continue reporting that disease status (based on labs / clinical assessments) until there is evidence of relapse / progression.
Example 1: A recipient with neuroblastoma is not in complete remission prior to transplant, in the 100-day reporting period the recipient receives a tandem transplant. Between HCT 1 and HCT 2 the only disease assessment performed was a clinical evaluation. In this case either option would be appropriate to answer for the current disease status: “Not evaluated” or “Not in complete remission (NCR)” and “no disease detected but incomplete evaluation to establish CR.”. However, we want to ensure the best response and the current disease status are consistent.

Example 2: A recipient with neuroblastoma is in complete remission prior to transplant, in the 100-day reporting period the recipient receives a tandem transplant. Between HCT 1 and HCT 2 the only disease assessment performed was a clinical evaluation in which the clinician did not mention progressive or relapsed disease. In this case “Complete remission (CR)” should be reported for the current disease status.

Question 125: Specify disease status if not in complete remission
Disease status criteria are generally based upon clinical assessment confirming ongoing presence or absence of disease. However, there are also situations in which an evaluation may have been performed but be incomplete and not have all testing required in order to meet the criteria for reporting complete remission (CR).

For recipients Not in complete remission, indicate whether clinical evidence of disease persisted on disease-specific assessments within the reporting period. If all assessments have shown resolution of disease, but not all assessments required to report complete remission have been completed, indicate No disease detected but incomplete evaluation to establish CR. This option is also appropriate for scenarios in which the recipient has not previously achieved a post-HCT CR but does not have any disease assessments performed within the reporting period. Indicate Disease detected if disease persists by any method of radiological or clinical assessment; persistence of abnormalities by molecular, cytogenetic, or flow cytometry assessments does not constitute “disease detected.”

Example 1: A recipient with multiple myeloma goes to transplant in VGPR, without a bone marrow showing < 5% blasts completed prior to transplant. Post-transplant serum and urine electrophoreses and immunofixations are negative. However, no bone marrow biopsy is performed within the 100-day reporting period. In this case, “not in complete remission” should be selected for question the current disease status, and “no disease detected by incomplete evaluation to establish CR” for the specify disease status if not in complete remission question.

Example 2: A recipient with AML goes to transplant in primary induction failure. Post-transplant, they recover their counts, but had circulating blasts noted on differential. They expire due to persistent disease with their last CBC performed on their date of death showing circulating blasts. In this case, “not in complete
“remission” should be selected for the current disease status, and “disease detected” for the specify disease status if not in complete remission question.

Example 3: Similar to example 2, a recipient with AML goes to transplant in primary induction failure. They expire on D+11 due to infection and had not engrafted as of that date. Their last CBC showed a WBC of 0.5 x 10⁹/L with no blasts detected on their differential. A bone marrow biopsy was not performed between transplant and the date of death. In this case, “not in complete remission” should be selected for the current disease status, and “no disease detected by incomplete evaluation to establish CR” for the specify disease status if not in complete remission question.

Questions 126 – 127: Date of assessment of current disease status
Report the date of latest clinical / hematologic assessment for the current disease status using the guidelines below:

- If the current disease status is Complete remission, report the date of the most disease specific clinical / hematologic assessment performed within approximately 30 days of the contact date.
- If the current disease status is Not in complete remission – disease detected, report the most recent clinical / hematologic assessment performed in the reporting period that detects disease.
- If the current disease status is Not in complete remission – no disease detected but incomplete evaluation to establish CR, report the last clinical / hematologic disease assessment performed in the reporting period.
- If there are no disease-specific assessments within the reporting period, report the latest assessment in which the recipient was clinically assessed by a physician or midlevel clinician. In this scenario, this date does not need to be consistent with the disease status reported in question 124 – 125.

Use known even if only approximate date is known, then refer to General Instructions, General Guidelines for Completing Forms, for information about reporting partial or unknown dates.

Example 1: The current disease status for a recipient with non-Hodgkin’s lymphoma is “complete remission.” A PET scan was performed 3 months prior to the contact date showing no evidence of disease and a physician’s exam was performed on the contact date. In this case, the physician’s exam performed on the contact date should be reported as the current disease assessment date since this is the most disease specific clinical / hematologic assessment performed within 30 days of the contact date.

Example 2: For a recipient with neuroblastoma, the current disease status is “not in complete remission – disease detected” since disease was still present on the last PET scan. The PET scan was performed 7 months prior to the contact date and a physician’s exam was performed on the contact date – disease cannot be detected by
the physician’s exam. The date of the PET scan should be reported as the current disease assessment date since this is the most disease specific clinical / hematologic assessment showing evidence of disease.

**Example 3**: The bone marrow biopsy performed for a recipient with AML still showed > 5% blasts in the bone marrow and therefore, the current disease status is reported as “not in complete remission – disease detected.” The bone marrow biopsy was performed 6 months prior to the contact date and a CBC was performed 2 weeks prior to the contact date – the CBC showed > 5% blasts in the blood. In this scenario, the current disease assessment date should be reported as the date of the CBC as this is the most recent disease specific clinical / hematologic assessment showing evidence of disease.

**Example 4**: A recipient with multiple myeloma had a bone marrow biopsy performed 2 weeks prior to the contact date which showed < 5% plasma cells; however, the last set of myeloma labs performed in the prior reporting period still showed evidence of disease; these labs were not repeated in the current reporting period. On the contact date, a physician’s exam was performed. The current disease status is “not in CR – no disease detected but incomplete evaluation to establish CR” and the current disease assessment date should be reported as the date of the physician’s exam as this is the last clinical / hematologic assessment performed in the reporting period.

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**Signature Lines**  
The FormsNet3SM application will automatically populate the signature data fields, including name and email address of person completing the form and date upon submission of the form.